Case study: *L. monocytogenes* in cold-smoked salmon

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The objective of this case study is to assess the risk of invasive listeriosis from consumption of cold-smoked salmon in France. The process of interest lays from the end of the production line in the factory, when the cold-smoked salmon is vacuum-packed, to the consumption.

The data and the model are adapted to illustrate the use of mc2d: the results will not and *should not* be interpreted as an assessment of the actual risk of listeriosis from consumption of cold-smoked salmon. Interested readers could refer to [3] and [2] for a complete risk assessment on that issue.

The model will be developed in a first section, without considering variability or uncertainty (deterministic model). Variability will then be introduced in a second section, and a last section will consider variability and a part of the data uncertainty.

1 The Model

In this section, no variability nor uncertainty is considered. We assess the final level of *L. monocytogenes* in the product, the exposure and the risk of invasive listeriosis for an “average” individual of the “healthy” French population\(^1\).

During the logistic, the retail and the home step, a bacterial growth is modeled considering i) the fluctuating temperature during the various stages and; ii) the bacterial competition with the food flora. We use the models developed and/or used in [3]. The data are adapted from [3] and [1]:

- The DMS model predicts the bacterial growth during a stage of duration \(d\), when the temperature is fluctuating, with an intra-stage average temperature \(m_T\) and an intra-stage standard deviation of the temperature \(s_T\). It is written:

\[
N_1 = \min \left( N_0 + \frac{\mu_{ref} \ln(10)}{\ln(10)} \times d \times \frac{s_T^2 + (m_T - T_{min})^2}{(T_{ref} - T_{min})^2}, N_{max} \right) \tag{1}
\]

if \(m_T > T_{min}\), with \(N_1\) the \(\log_{10}\) concentration of bacteria (\(\log_{10}\) (CFU/g)) in the product at the end of the stage, \(N_0\) the \(\log_{10}\) concentration of bacteria (\(\log_{10}\) (CFU/g)) in the product at the beginning of the stage, \(\mu_{ref}\) the specific growth rate (day\(^{-1}\)) at a reference temperature \(T_{ref}\) (°C), \(T_{min}\) the minimal temperature (°C) of growth and \(N_{max}\) the maximum achievable concentration in the product (\(\log_{10}\) (CFU/g)). If \(m_T \leq T_{min}\), \(N_1 = N_0\).

- We will use \(T_{ref} = 25^\circ\text{C}\). We have in this section \(N_{max} = 7.27 \log_{10}\text{(CFU/g)}\);

- The model for *L. monocytogenes* uses \(\mu_{ref,Lm} = 6.2\text{ day}^{-1}\) and \(T_{min,Lm} = -2.9^\circ\text{C}\);

- The same model is used for the food flora, with \(\mu_{ref,ff} = 4.1\text{ day}^{-1}\) and \(T_{min,ff} = -4.5^\circ\text{C}\);

- The growth model for the bacterial competition consider the Jameson effect, i.e. consider that the bacterial growth of *L. monocytogenes* and the growth of the food flora are stopped as soon as one population reaches \(N_{max}\).

\(^1\)It makes little sense, but it will help us introducing smoothly the model.
In practice, one will evaluate $d_{Lm}$ and $d_{ff}$, the time needed for *L. monocytogenes* or the food flora to reach $N_{max}$, respectively, and model a growth for the given stage during an effective duration of $\min(d, d_{Lm}, d_{ff})$. The time needed to reach $N_{max}$ is evaluated by inverting (1):

$$d_{N_1=N_{max}} = \left( N_{max} - N_0 \right) \times \frac{\ln(10)}{\mu_{ref}} \times \frac{(T_{ref} - T_{min})^2}{\left( s_T^2 + (m_T - T_{min})^2 \right)}$$

The other assumptions are:

- A cold-smoked salmon package is homogeneously contaminated with *L. monocytogenes* at the end of the production at a level of 0.1 CFU/g;
- The food flora level at the end of the production is $10^{2.78}$ CFU/g;
- The time-temperature profile is:
  - 1.1 days at an average temperature of 3.2°C from the factory to the retail (logistic step), with an intra-stage standard deviation of the temperature of 2.1°C;
  - 4.7 days at an average temperature of 5.5°C at retail with an intra-stage standard deviation of the temperature of 1.0°C;
  - 4.3 days at an average temperature of 8.2°C in the consumer’s home with an intra-stage standard deviation of the temperature of 2.0°C;
- An healthy, non elderly, non pregnant individual eats 35g of this product;
- The individual dose-response model for this population is a one hit model
  $$\Pr(\text{Illness} \mid D) = 1 - (1 - r)^D$$
  with $r = 4.7 \times 10^{-14}$ for an individual from this healthy sub-population. The populational dose-response that evaluates the mean risk for a population exposed to food where the number of bacteria follows a Poisson distribution of mean parameter $D$ is the exponential dose-response
  $$\Pr(\text{Illness} \mid D) = 1 - \exp(r \times D)$$

The question is “What is the risk for this ‘average’ individual?”. One way to write this model is as following:

```r
> Nmax <- 7.3
> murefLm <- 6.2; TminLm <- -2.9
> mureffF <- 4.1; TminFF <- -4.5
> Lm0 <- log10(1); FF0 <- 2.78
> d1 <- 1.1; mT1 <- 3.2; sdT1 <- 2.1
> d2 <- 4.7; mT2 <- 5.5; sdT2 <- 1.0
> d3 <- 4.3; mT3 <- 8.2; sdT3 <- 2.0
> conso <- 35
> r <- 4.7e-14
> modGrowth <- function(duration, mTemp, sdTemp,
+   NOLm, murefLm, TminLm,
+   NOFF, mureffF, TminFF,
+   Nmax, Tref=25) {
+     NOLm <- pmin(NOLm, Nmax)
+     NOFF <- pmin(NOFF, Nmax)
+     dLm <- (Nmax-NOLm) * log(10)/murefLm * (Tref-TminLm)^2 / (sdTemp^2 + (mTemp-TminLm)^2)
+     dFF <- (Nmax-NOFF) * log(10)/mureffF * (Tref-TminFF)^2 / (sdTemp^2 + (mTemp-TminFF)^2)
+     return(dLm, dFF)
+ }
```
modGrowth is a convenient function for the growth model. Within this function \(dLm\) is the time needed for \(L.\) monocytogenes to reach \(Nmax\), \(dFF\) is the time needed for the food flora to reach \(Nmax\) and, \(realDuration\) is the effective time of growth during the stage. Note that:

- this function is “vectorized”, meaning that it can deal with a vector for any of its parameters, returning consequently a vector. This is a strength of R, notably for Monte-Carlo simulations, but it requests a bit of knowledge on the way to code the functions. As an example: \(\text{pmin}\), a function that takes one or more vectors as arguments and return a single vector giving the “parallel” minima of the vectors is used instead of the more classical function \(\text{min}\) function, that would return the maximum or minimum of all the values. Another example is the use of the \(\text{ifelse}\) instead of \(\text{if}\);

- it is also written to handle all specific cases that could occur in the Monte-Carlo simulation, such as \(N_0 \geq N_{max}\) or \(m_T \leq T_{min}\) or both, for any or both bacterial populations.

\(x1\), \(x2\) and \(x3\) are the bacterial concentrations at the end of the logistic, the retail and the home step, respectively.
2 Including Variability

We now specify now some variability distributions for some inputs, following [1] and [3]. We first have to call the needed libraries, and define the desired number of iterations:

> library(fitdistrplus)
> library(mc2d)
> ndvar(10001)

[1] 10001

2.1 Specifying Variability Distribution

2.1.1 Initial Contamination

For the initial contamination levels in *L. monocytogenes*, we have a set of 62 enumeration data from a representative sample of packages of cold smoked salmon positive in detection: 43 samples have less than 0.2 CFU/g, 7 samples have 0.2 CFU/g, 4 samples have 0.4 CFU/g, 2 samples have 0.6 CFU/g, and the other values are 0.3, 1.0, 1.6, 2.4, 5.4 and 7.0 CFU/g [3]. We will use the fitdistrplus package to fit a normal distribution on the log_{10} of these values, taking into account the censored values. Using the fitted parameters, we model thereafter these initial concentrations in contaminated packages through a normal distribution truncated^2 on [−2, ∞) log_{10} (CFU/g).

For the food flora, we use the distribution proposed by [1], \( N_{ff} \sim N(2.78, 1.14) \).

> dataC <- data.frame(
+   left = c(rep(NA, 43), rep(0.2, 7), 0.3, rep(0.4, 4), 1, 1.6, 6, 2.4, 5.4, 7),
+   right = c(rep(0.2, 43), rep(0.2, 7), 0.3, rep(0.4, 4), 1, 1.6, 6, 2.4, 5.4, 7)
+ )
> fit <- fitdistcens(log10(dataC), "norm")
> fit

Fitting of the distribution ' norm ' on censored data by maximum likelihood

Parameters:
  estimate
  mean  -1.117
  sd  0.764
Fixed parameters:
  data frame with 0 columns and 0 rows

> Lm0V <- mcstoc(rnorm, mean = fit$est["mean"], sd = fit$est["sd"], rtrunc=TRUE, linf=-2)
> FF0V <- mcstoc(rnorm, mean=2.78, sd=1.14)

Note that, by default, the type of alea that is modeled is “variability” (type="V").

2.1.2 Growth Parameters

Distributions are derived from [1]:

- \( N_{max} \) follows a normal distribution with mean 7.27 log_{10} CFU/g and standard deviation 0.86 log_{10} CFU/g;
- The specific growth rate at the reference temperature of 25°C for *L. monocytogenes* follows a normal distribution with mean 6.24 day\(^{-1}\) and standard deviation 0.75 day\(^{-1}\) truncated on [0, ∞). The minimal growth temperature follows a normal distribution with mean -2.86°C and standard deviation 1.93°C;
- The specific growth rate at the reference temperature of 25°C for the food flora follows a normal distribution with mean 4.12 day\(^{-1}\) and standard deviation 1.97 day\(^{-1}\) truncated on [0, ∞). The minimal growth temperature follows a normal distribution with mean -4.52°C and standard deviation 7.6°C.

\(^2\)so that at least one CFU is included in one 100g package
Table 1: Time Temperature Profiles

<table>
<thead>
<tr>
<th>Stage</th>
<th>Mean Temperature (°C)</th>
<th>Intra-Stage Variance of T (°C)</th>
<th>time (days)</th>
</tr>
</thead>
<tbody>
<tr>
<td>logistic</td>
<td>normal(3.2, 2.2) truncated on [3:25]</td>
<td>Γ(1.16, 4.61)</td>
<td>Exponential(1.1)</td>
</tr>
<tr>
<td>retail</td>
<td>normal(5.5, 2.2) truncated on [3:25]</td>
<td>Γ(0.65, 2.09)</td>
<td>Exponential(4.7)</td>
</tr>
<tr>
<td>consumer</td>
<td>normal(8.2, 3.8) truncated on [3:25]</td>
<td>Γ(0.35, 19.7)</td>
<td>Exponential(4.3)</td>
</tr>
</tbody>
</table>

> NmaxV <- mcstoc(rnorm, mean=7.27, sd = 0.86)
> murefLmV <- mcstoc(rnorm, mean = 6.24, sd = 0.75, rtrunc=TRUE, linf=0)
> TminLmV <- mcstoc(rnorm, mean = -2.86, sd = 1.93)
> murefFFV <- mcstoc(rnorm, mean = 4.12, sd = 1.97, rtrunc=TRUE, linf=0)
> TminFFV <- mcstoc(rnorm, mean = -4.52, sd = 7.66)

2.1.3 Time-Temperature Profiles

The time temperature profiles in the three steps are modelled using the distribution provided in the table 1 (adapted from [3] from representative data from France)\(^3\). We assume a shelf life of 28 days. A simple way to model this shelf life will be to have \(d_1 + d_2 + d_3 \leq 28\) days, with \(d_1\) the duration of the logistic stage, \(d_2\) the duration of the retail stage and \(d_3\) the duration of the consumer stage\(^4\):

> d1V <- mcstoc(rexp, rate = 1/1.1)
> mT1V <- mcstoc(rnorm, mean = 3.2, sd = 2.2, rtrunc = TRUE, linf = -3, lsup = 25)
> sdT1V <- sqrt(mcstoc(rgamma, shape = 1.16, scale=4.61))
> d2V <- mcstoc(rexp, rate = 1/4.7, rtrunc=TRUE, lsup=28-d1V)
> mT2V <- mcstoc(rnorm, mean = 5.5, sd = 2.2, rtrunc = TRUE, linf = -3, lsup = 25)
> sdT2V <- sqrt(mcstoc(rgamma, shape = 0.65, scale=2.09))
> d3V <- mcstoc(rexp, rate = 1/4.3, rtrunc=TRUE, lsup=28-(d1V+d2V))
> mT3V <- mcstoc(rnorm, mean = 8.2, sd = 3.8, rtrunc = TRUE, linf = -3, lsup = 25)
> sdT3V <- sqrt(mcstoc(rgamma, shape = 0.35, scale=19.7))

2.1.4 Serving Size

As for the serving size, we consider, from observed data, a discrete empirical distribution with values [3]: \(V = \{10, 12, 19, 20, 30, 34, 40, 50, 60, 67.5, 80, 100, 250\} \) grams, observed \(F = \{11, 1, 1, 29, 12, 1, 41, 4, 4, 1, 4, 1, 1\} \) time, respectively.

> consoV <- mcstoc(remipiricalD,
+ values = c(10, 12, 19, 20, 30, 34, 40, 50, 60, 67.5, 80, 100, 250),
+ prob = c(11, 1, 1, 29, 12, 1, 41, 4, 4, 1, 4, 1, 1))

2.2 Applying the Model

The model may then be evaluated straightforwardly:

> r <- mcdata(4.7e-14, type = "0")
> x1V <- modGrowth(d1V, mT1V, sdT1V,
+ Lm0V, murefLmV, TminLmV,
+ FF0V, murefFFV, TminFFV,
+ NmaxV)
> x2V <- modGrowth(d2V, mT2V, sdT2V,
+ x1V$Lm, murefLmV, TminLmV,
+ x1V$FF, murefFFV, TminFFV,
+ NmaxV)

---

\(^3\)Γ is the Gamma distribution parameterized as Γ(shape, scale). The Exponential(x) distribution is the exponential distribution with mean x.

\(^4\)See the code for a way to model this shelf life using truncated distributions.
> x3V <- modGrowth(d3V, mT3V, sdT3V,
+       x2V$xLm, murefLmV, TminLmV,
+       x2V$xFF, murefFFV, TminFFV,
+       NmaxV)
> contaV <- 10^x3V$xLm
> expoV <- consoV * contaV
> riskV <- 1 - exp(-r * expoV )
> Lm1 <- mc(Lm0V, FF0V, NmaxV, murefLmV, TminLmV, murefFFV, TminFFV,
+       d1V, mT1V, sdT1V, d2V, mT2V, sdT2V, d3V, mT3V, sdT3V,
+       consoV, r, contaV, expoV, riskV)

```
node mode nsv nsu nva variate min mean median max Nas type outm
1  Lm0V numeric 10001 1 1 1 -2.00e+00 -9.30e-01 -9.88e-01 1.76e+00 0 V each
2  FF0V numeric 10001 1 1 1 -1.28e+00 2.78e+00 2.78e+00 6.85e+00 0 V each
3  NmaxV numeric 10001 1 1 1 3.97e+00 7.26e+00 7.27e+00 1.06e+01 0 V each
4  murefLmV numeric 10001 1 1 1 2.85e+00 6.24e+00 6.25e+00 9.25e+00 0 V each
5  TminLmV numeric 10001 1 1 1 -1.01e+01 -2.83e+00 -2.85e+00 3.69e+00 0 V each
6  murefFFV numeric 10001 1 1 1 1.31e-02 4.19e+00 4.17e+00 1.13e+01 0 V each
7  TminFFV numeric 10001 1 1 1 -3.51e+01 -4.52e+00 -4.46e+00 2.66e+01 0 V each
8  d1V numeric 10001 1 1 1 5.36e-05 1.10e+00 7.69e-01 9.69e+00 0 V each
9  mT1V numeric 10001 1 1 1 -2.98e+00 3.20e+00 3.15e+00 1.14e+01 0 V each
10 sdT1V numeric 10001 1 1 1 3.37e-02 2.08e+00 1.96e+00 6.57e+00 0 V each
11  d2V numeric 10001 1 1 1 1.70e-03 4.69e+00 3.29e+00 2.67e+01 0 V each
12  mT2V numeric 10001 1 1 1 -2.53e+00 5.53e+00 5.50e+00 1.35e+01 0 V each
13 sdT2V numeric 10001 1 1 1 1.60e-03 9.66e-01 8.63e-01 4.76e+00 0 V each
14  d3V numeric 10001 1 1 1 3.84e-04 4.09e+00 2.89e+00 2.53e+01 0 V each
15  mT3V numeric 10001 1 1 1 -2.98e+00 8.24e+00 8.21e+00 2.26e+01 0 V each
16 sdT3V numeric 10001 1 1 1 1.31e-06 1.91e+00 1.41e+00 1.16e+01 0 V each
17  consoV numeric 10001 1 1 1 1.00e+01 3.55e+01 4.00e+01 2.50e+02 0 V each
18  r numeric 1 1 1 1 4.70e-14 4.70e-14 4.70e-14 4.70e-14 0 0 each
19 contaV numeric 10001 1 1 1 1.16e-02 4.09e+06 2.22e+01 5.04e+09 0 V each
20 expoV numeric 10001 1 1 1 1.70e-01 1.18e+08 6.85e+02 1.01e+11 0 V each
21 riskV numeric 10001 1 1 1 7.99e-15 5.55e-06 3.22e-11 4.73e-03 0 V each
```
> sLm1 <- mc(contaV=Lm1$contaV, expoV=Lm1$expoV, riskV=Lm1$riskV)
> summary(sLm1, probs = c(0, 0.5, 0.75, 0.95, 1))

contaV :

<table>
<thead>
<tr>
<th>mean</th>
<th>sd</th>
<th>Min</th>
<th>50%</th>
<th>75%</th>
<th>95%</th>
<th>Max</th>
<th>nsv</th>
<th>Na's</th>
</tr>
</thead>
<tbody>
<tr>
<td>4092015</td>
<td>76520808</td>
<td>0.0116</td>
<td>22.2</td>
<td>867</td>
<td>1962680</td>
<td>5.04e+09</td>
<td>10001</td>
<td>0</td>
</tr>
</tbody>
</table>

expoV :

<table>
<thead>
<tr>
<th>mean</th>
<th>sd</th>
<th>Min</th>
<th>50%</th>
<th>75%</th>
<th>95%</th>
<th>Max</th>
<th>nsv</th>
<th>Na's</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.18e+08</td>
<td>1.88e+09</td>
<td>0.17</td>
<td>685</td>
<td>27251</td>
<td>60077024</td>
<td>1.01e+11</td>
<td>10001</td>
<td>0</td>
</tr>
</tbody>
</table>

riskV :

<table>
<thead>
<tr>
<th>mean</th>
<th>sd</th>
<th>Min</th>
<th>50%</th>
<th>75%</th>
<th>95%</th>
<th>Max</th>
<th>nsv</th>
<th>Na's</th>
</tr>
</thead>
<tbody>
<tr>
<td>5.55e-06</td>
<td>8.82e-05</td>
<td>7.99e-15</td>
<td>3.22e-11</td>
<td>1.28e-09</td>
<td>2.82e-06</td>
<td>0.00473</td>
<td>10001</td>
<td>0</td>
</tr>
</tbody>
</table>

Lm1 is a `mc` object that contains all the parameters and outputs. We extract some of these outputs in `sLm1` to provide a short summary.

Lm1 is a `mc` object that contains all the parameters and outputs. We extract some of these outputs in `sLm1` to provide a short summary.
2.3 Final Estimate

If 6.5% of cold-smoked salmon package are contaminated, if 49,090,000 Frenchs are part of the “non susceptible” population and if, on average, those people consume some smoked salmon 6.4 times per year, the expected number of cases of listeriosis from consumption of cold smoked salmon in this population is estimated through:

```r
> meanRisk <- mcapply(riskV,"var","mean")
> expectedN <- round(0.065 * unmc(meanRisk) * 6.4 * 49090000)
> expectedN
[1] 113
```

3 Including (a Part of the) Uncertainty

We eventually include both variability and uncertainty in the model. For this example, we will only consider the uncertainty linked to the initial contamination, the growth parameters and the prevalence.

3.1 Specifying Uncertainty

3.1.1 Initial Contamination

The uncertainty surrounding the initial contamination levels of the *L. monocytogenes* will be modeled using a bootstrap procedure, obtained straightforwardly with the help of the *fitdistrplus* package and its *bootdistcens* function. Before this, we define the number of iterations needed in the uncertainty dimension.

```r
> ndunc(101)
[1] 101
```

```r
> bootLm0 <- bootdistcens(fit, niter=ndunc())
> MLm0 <- mcdata(bootLm0$est$mean,type="U")
> SLm0 <- mcdata(bootLm0$est$sd,type="U")
> Lm0VU <- mcstoc(rnorm, type="VU", mean=MLm0, sd=SLm0, rtrunc=TRUE, linf=-2)
```

In order to consider uncertainty for the food flora initial contamination, we have, from [1], a set of uncertain hyperparameters, $M_{N_{0_{ff}}}$ and $\sigma_{N_{0_{ff}}}$, that are used as parameters for the uncertain and variable parameter $N_{0_{ff}}$:

\[
N_{0_{ff}} \sim N(M_{N_{0_{ff}}}, \sigma_{N_{0_{ff}}})
\]

\[
M_{N_{0_{ff}}} \sim N(2.78, 0.265)
\]

\[
\ln(\sigma_{N_{0_{ff}}}) \sim N(0.114, 0.172)
\]

This hierarchical simulation is written with *mc2d*:

```r
> MLm0FF <- mcstoc(rnorm, type="U", mean=2.78, sd=0.265)
> SLm0FF <- mcstoc(rlnorm, type="U", meanlog=0.114, sdlog=0.172)
> FF0VU <- mcstoc(rnorm, type="VU", mean=MLm0FF, sd=SLm0FF)
```
3.1.2 Growth Parameters

The uncertainty around $\mu_{ref, Lm}$, $T_{min, Lm}$, $\mu_{ref, ff}$, $T_{min, ff}$ and $N_{max}$ are modeled similarly through the specification of hyperparameters [1]5:

$$\mu_{ref, Lm} \sim N(M_{\mu_{ref, Lm}}, \sigma_{\mu_{ref, Lm}})$$

$$M_{\mu_{ref, Lm}} \sim \Gamma(\text{shape}: 69.7, \text{scale}: 0.0896)$$

$$\ln(\sigma_{\mu_{ref, Lm}}) \sim N(1.03, 0.191)$$

$$T_{min, Lm} \sim N(M_{T_{min, Lm}}, \sigma_{T_{min, Lm}})$$

$$M_{T_{min, Lm}} \sim N(-2.86, 0.459)$$

$$\ln(\sigma_{T_{min, Lm}}) \sim N(0.638, 0.208)$$

$$\mu_{ref, ff} \sim N(M_{\mu_{ref, ff}}, \sigma_{\mu_{ref, ff}})$$

$$M_{\mu_{ref, ff}} \sim \Gamma(\text{shape}: 32.5, \text{scale}: 0.127)$$

$$\ln(\sigma_{\mu_{ref, ff}}) \sim N(-0.656, 0.221)$$

$$T_{min, ff} \sim N(M_{T_{min, ff}}, \sigma_{T_{min, ff}})$$

$$M_{T_{min, ff}} \sim N(-4.52, 1.23)$$

$$\ln(\sigma_{T_{min, ff}}) \sim N(2.00, 0.257)$$

$$N_{max} \sim N(M_{N_{max}}, \sigma_{N_{max}})$$

$$M_{N_{max}} \sim N(7.27, 0.276)$$

$$\ln(\sigma_{N_{max}}) \sim N(-0.172, 0.218)$$

with $\mu_{ref} > 0$ and $T_{min} < 25$. We simply translated the preceding distributions:

```r
> MmurefLm <- mcstoc(rgamma, type="U", shape=69.7, scale=0.0896)
> SmurefLm <- mcstoc(rlnorm, type="U", meanlog = 1.03, sdlog = 0.191)
> murefLmVU <- mcstoc(rnorm, type="VU", mean=MmurefLm, sd=SmurefLm, rtrunc=TRUE, linf=0)
> MTminLm <- mcstoc(rnorm, type="U", mean=-2.86, sd=0.459)
> STminLm <- mcstoc(rlnorm, type="U", meanlog = 0.638, sdlog = 0.208)
> TminLmVU <- mcstoc(rnorm, type="VU", mean = MTminLm, sd = STminLm, rtrunc=TRUE, lsup=25)
> MmurefFF <- mcstoc(rgamma, type="U", shape=32.5, scale=.127)
> SmurefFF <- mcstoc(rlnorm, type="U", meanlog = -.656, sdlog = 0.221)
> murefFFVU <- mcstoc(rnorm, type="VU", mean=MmurefFF, sd=SmurefFF, rtrunc=TRUE, linf=0)
> MTminFF <- mcstoc(rnorm, type="U", mean=-4.52, sd=1.23)
> STminFF <- mcstoc(rlnorm, type="U", meanlog = 2.00, sdlog = 0.257)
> TminFFVU <- mcstoc(rnorm, type="VU", mean = MTminFF, sd = STminFF, rtrunc=TRUE, lsup=25)
> MNmax <- mcstoc(rnorm, type="U", mean=7.27, sd=0.276)
> SNmax <- mcstoc(rlnorm, type="U", meanlog = -0.172, sdlog = 0.218)
> NmaxVU <- mcstoc(rnorm, type="VU", mean = MNmax, sd = SNmax)
```

3.1.3 Prevalence

The prevalence level of contaminated cold-smoked salmon packages (6.5%) was estimated from 41 positive packages out of 626 tested [3]. We assume a sensitivity and a specificity of the method of 100%. We model the data

5Note that there was a typo in [1] that lead to an error in [3]: the standard-error for $\ln(\sigma_{\mu_{ref, Lm}})$ is 1.03 and not −1.03 as written in [1]. We will use here the correct value.
uncertainty around the true prevalence of contaminated package using a bayesian reasoning, with a Beta(1, 1) distribution as a prior. The number of expected cases may be estimated using:

```R
> prevU <- mcstoc(rbeta,type="U", shape1=41+1, shape2=626-41+1)
```

### 3.2 Applying the Model

Applying the model is just a copy-paste from the previous version (+ we change the name of the parameters).

```R
> x1VU <- modGrowth(d1V, mT1V, sdT1V,
+    Lm0VU, murefLmVU, TminLmVU,
+    FFOVU, mureffFFVU, TminFFVU,
+    NmaxVU)
> x2VU <- modGrowth(d2V, mT2V, sdT2V,
+    x1VU$xLm, murefLmVU, TminLmVU,
+    x1VU$xFF, mureffFFVU, TminFFVU,
+    NmaxVU)
> x3VU <- modGrowth(d3V, mT3V, sdT3V,
+    x2VU$xLm, murefLmVU, TminLmVU,
+    x2VU$xFF, mureffFFVU, TminFFVU,
+    NmaxVU)
> contaVU <-10^x3VU$xLm
> expoVU <- consoV * contaVU
> riskVU <- 1 - exp(-r * expoVU)
> Lm2 <- mc(Lm0VU, FF0VU, NmaxVU, murefLmVU, TminLmVU, mureffFFVU, TminFFVU,
+    d1V, mT1V, sdT1V, d2V, mT2V, sdT2V, d3V, mT3V, sdT3V,
+    consoV, r, contaVU, expoVU, riskVU)
> Lm2
```

```R
node mode nsv nsu nva variate min mean median max Nas type outm
1 Lm0VU numeric 10001 101 1 1 -2.00e+00 -9.37e-01 -9.93e-01 3.68e+00 0 VU each
2 FFOVU numeric 10001 101 1 1 -4.82e+00 2.76e+00 2.76e+00 9.58e+00 0 VU each
3 NmaxVU numeric 10001 101 1 1 2.11e+00 7.28e+00 7.28e+00 1.27e+11 0 VU each
4 murefLmVU numeric 10001 101 1 1 1.00e-04 6.45e+00 6.36e+00 2.43e+00 0 VU each
5 TminLmVU numeric 10001 101 1 1 -1.44e+01 -2.83e+00 -2.84e+00 8.70e+00 0 VU each
6 mureffFFVU numeric 10001 101 1 1 4.81e-03 4.21e+00 4.21e+00 8.24e+00 0 VU each
7 TminFFVU numeric 10001 101 1 1 -5.90e+01 -4.39e+00 -4.35e+00 2.50e+00 0 VU each
8 d1V numeric 10001 1 1 1 5.36e-05 1.10e+00 7.69e-01 9.69e+00 0 V each
9 mT1V numeric 10001 1 1 1 -2.98e+00 3.20e+00 3.15e+00 1.14e+00 0 V each
10 sdT1V numeric 10001 1 1 1 3.37e-02 2.08e+00 1.96e+00 6.57e+00 0 V each
11 d2V numeric 10001 1 1 1 1.70e-03 4.69e+00 3.29e+00 2.67e+01 0 V each
12 mT2V numeric 10001 1 1 1 -2.53e+00 5.53e+00 5.50e+00 1.35e+01 0 V each
13 sdT2V numeric 10001 1 1 1 1.60e-03 9.66e-01 8.63e-01 4.76e+00 0 V each
14 d3V numeric 10001 1 1 1 3.84e-04 4.09e+00 2.89e+00 2.53e+00 0 V each
15 mT3V numeric 10001 1 1 1 -2.98e+00 8.24e+00 8.21e+00 2.62e+01 0 V each
16 sdT3V numeric 10001 1 1 1 1.31e-06 1.91e+00 1.41e+00 1.16e+01 0 V each
17 consoV numeric 10001 1 1 1 1.00e+01 3.55e+01 4.00e+01 2.50e+02 0 V each
18 r numeric 1 1 1 4.70e-14 4.70e-14 4.70e-14 4.70e-14 0 V each
19 contaVU numeric 10001 101 1 1 1.01e-02 1.24e+07 1.71e+01 5.74e+11 0 VU each
20 expoVU numeric 10001 101 1 1 1.01e-01 4.53e+08 5.22e+02 2.87e+13 0 VU each
21 riskVU numeric 10001 101 1 1 4.77e-15 2.03e-05 2.46e-11 7.41e-01 0 VU each
```

```R
> sLm2 <- mc(contaVU=Lm2$contaVU, expoVU=Lm2$expoVU, riskVU=Lm2$riskVU)
> summary(sLm2, probs = c(0, 0.5, 0.75, 0.95, 1))
```
The summary provides the estimate of the mean, the standard deviation, the minimum, the median... and a 95% credible interval. The estimate is the median of the 101 values obtained in the uncertainty dimension. The credible interval lays between the 2.5th and the 97.5th percentiles obtained in the uncertainty dimension.

### 3.3 Final Estimate

The uncertainty around the number of expected cases is estimated using:

```r
> meanRiskU <- mcapply(riskVU, "var", mean)
> expectedNU <- round(prevU * meanRiskU * 6.4 * 49090000)
> summary(expectedNU)
```

<table>
<thead>
<tr>
<th>node</th>
<th>median</th>
<th>mean</th>
<th>2.5%</th>
<th>97.5%</th>
</tr>
</thead>
<tbody>
<tr>
<td>NoVar</td>
<td>267</td>
<td>423</td>
<td>38</td>
<td>1954</td>
</tr>
</tbody>
</table>

This is an estimate of the uncertainty around the number of cases linked to the uncertainty around the initial contamination, the bacterial growth parameter and the sampling uncertainty for positive packages. A lot of other uncertainties exist but are not considered here, notably the uncertainty around the dose-response model and parameters. See [3, 2] for a complete analysis. The study of the model through a Tornado chart in the variability dimension leads to the Figure 1. It suggests a big impact of the growth rate of *L. monocytogenes*, of the storage duration during the consumer step, and of the initial level of *L. monocytogenes*. The Tornado chart in the uncertainty dimension leads to the Figure 2 and suggests the impact of the uncertainty around $N_{\text{max}}$ on the mean risk, and thus the expected number of cases.

```r
> torn <- tornado(Lm2)
> torn
```

Spearman's rho statistic
Output: riskVU
$\text{riskVU}$
Lm0VU  FFOVU  NmaxVU  murefLmVU  TminLmVU  murefFFVU  TminFFVU  d1V  mT1V  sdT1V  d2V  
median  0.303 -0.0823  0.0711  0.465 -0.237 -0.0292  0.1184  0.0447  0.0337  0.00419  0.277  
mean  0.298 -0.0888  0.0751  0.457 -0.238 -0.0306  0.1257  0.0440  0.0334  0.00361  0.277  
2.5%  0.202 -0.1567  0.0293  0.347 -0.342 -0.0568  0.0444  0.0245  0.0197 -0.01155  0.227  
97.5%  0.380 -0.0340  0.1382  0.548 -0.159 -0.00703  0.2246  0.0608  0.0488  0.01783  0.328  
mT2V  sdT2V  d3V  mT3V  sdT3V  consoV  contaVU  expoVU  
median  0.158  0.00741  0.406  0.260  0.0291  0.125  0.991  1  
mean  0.158  0.00714  0.407  0.259  0.0296  0.125  0.990  1  
2.5%  0.132 -0.00798  0.330  0.219  0.0165  0.105  0.986  1  
97.5%  0.187  0.02019  0.479  0.309  0.0418  0.148  0.994  1  
> tornunc <- tornadounc(Lm2, quant=.975)  
> tornunc  

Tornado on uncertainty  
Spearman's rho statistic  
Output: riskVU  
$riskVU  
mean Lm0VU  sd Lm0VU  97.5% Lm0VU  mean FFOVU  sd FFOVU  97.5% FFOVU  mean NmaxVU  sd NmaxVU  
mean riskVU  0.155  0.04478  0.0549 -0.0904 -0.170 -0.195  0.656  0.720  
mean riskVU  0.161 -0.00829  0.0139 -0.0186 -0.179 -0.152  0.509  0.796  
97.5% riskVU  0.186  0.17879  0.1750 -0.2009 -0.120 -0.228  0.735  0.242  
mean riskVU  0.912  0.409  0.286  0.439  0.0176 -0.0524  
mean riskVU  0.900  0.280  0.179  0.296  0.0736 -0.1391  
97.5% riskVU  0.552  0.639  0.428  0.639 -0.0901  0.0917  
97.5% riskVU  -0.0305 -0.1796  0.0659 -0.1505  0.159  0.003891  
97.5% riskVU  -0.0806 -0.0773  0.0337 -0.0649  0.140  0.000711  
97.5% riskVU  0.0446 -0.3025  0.0494 -0.2760  0.173  0.025172  
mean riskVU  0.0269  0.994  0.931  0.780  1.000  0.936  
mean riskVU  0.0111  0.928  0.967  0.571  0.938  1.000  
mean riskVU  0.0569  0.769  0.562  0.997  0.772  0.567  
mean riskVU  0.776  
mean riskVU  0.568  
97.5% riskVU  1.000  
> plot(torn)  
> plot(tornunc, stat="mean risk")  

As a conclusion, this example illustrates how predictive growth models may be implemented within mc2d . . .  

References  
Figure 1: Tornado chart for the *L. monocytogenes* example (Variability).

Figure 2: Tornado chart for the *L. monocytogenes* example (Uncertainty).