Package ‘qtlnet’

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Author  Elias Chaibub Neto <echaibub@hotmail.com> and Brian S. yandell <byandell@wisc.edu>
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Maintainer Brian S. Yandell <byandell@wisc.edu>
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**Acyclic graph example**

**Description**

We generate synthetic data (sample size 300) according to a DAG composed by 100 nodes and 107 edges (exactly as in Figure 1). Each phenotype node is affected by three QTLs, and we allow only additive genetic effects. The QTLs for each phenotype are randomly selected among 200 markers, with 10 markers unevenly distributed on each of 20 autosomes. We allowed different phenotypes to potentially share common QTLs. For each phenotype, the regression coefficients with other phenotypes are chosen uniformly between 0.5 and 1; QTL effects are chosen between 0.2 to 0.6; and residual standard deviations are chosen from 0.1 to 0.5. For each realization we apply the QDG algorithm to infer causal directions for the edges of the skeleton obtained by the PC-skeleton algorithm.

**Usage**

```r
data(acyclic)
```

**Details**

For cyclic graphs, the output of the `qdg` function computes the log-likelihood up to the normalization constant (un-normalized log-likelihood). We can use the un-normalized log-likelihood to compare cyclic graphs with reversed directions (since they have the same normalization constant). However we cannot compare cyclic and acyclic graphs.

**References**


**See Also**

`sim.cross`, `sim.geno`, `sim.map`, `skeleton`, `qdg`, `graph.qdg`, `generate.qtl.pheno`
Examples

```r
## Not run:
## This reproduces Figure 1 exactly.
set.seed(3456789)

tmp <- options(warn=-1)
acyclic.DG <- randomDAG(n = 100, prob = 2 / 99)

options(tmp)

## Simulate cross object using R/qtl routines.
n.ind <- 300
mymap <- sim.map(len=rep(100,20), n.mar=10, eq.spacing=FALSE, include.x=FALSE)
mycross <- sim.cross(map=mymap, n.ind=n.ind, type="f2")
summary(mycross)
mycross <- sim.geno(mycross,n.draws=1)

## Produce 100 QTL at three markers apiece.
acyclic.qtl <- generate.qtl.markers(cross=mycross,n.phe=100)

## Generate data from directed graph.
bp <- runif(100,0.5,1)
stdev <- runif(100,0.1,0.5)
bq <- matrix(0,100,3)
bq[,1] <- runif(100,0.2,0.4)
bq[,2] <- bq[,1]+0.1
bq[,3] <- bq[,2]+0.1

## Generate phenotypes.
acyclic.data <- generate.qtl.pheno("acyclic", cross = mycross, bp = bp, bq = bq, stdev = stdev, allqtl = acyclic.qtl$allqtl)

acyclic.qdg <- qdg(cross=acyclic.data, phenotype.names=paste("y",1:100,sep=""), marker.names=acyclic.qtl$markers, QTL=acyclic.qtl$allqtl, alpha=0.005, n.qdg.random.starts=1, skel.method="pcskel")
save(acyclic.DG, acyclic.qtl, acyclic.data, acyclic.qdg, file = "acyclic.RData", compress = TRUE)

## End(Not run)

data(acyclic)

dims <- dim(acyclic.data$pheno)
SuffStat <- list(C = cor(acyclic.data$pheno), n = dims[1])
pc <- skeleton(SuffStat, gaussCItest, p = dims[2], alpha = 0.005)
summary(pc)

summary(graph.qdg(acyclic.qdg))
```
bic.qtlnet

Pre-compute BIC values for qtlnet sampling.

Description

Pre-compute BIC values for qtlnet sampling to speed up MCMC sampling.

Usage

bic.qtlnet(cross, pheno.col, threshold, addcov = NULL, intcov = NULL,
max.parents = 3, parents, verbose = TRUE, ...)
bic.join(cross, pheno.col, ..., max.parents = 3)
data(Pscdbp.bic)

Arguments

cross Object of class cross. See read.cross.

pheno.col Phenotype identifiers from cross object. May be numeric, logical or character.

threshold Scalar or list of thresholds, one per each node.

addcov Additive covariates for each phenotype (NULL if not used). If entered as scalar or vector (same format as pheno.col), then the same addcov is used for all phenotypes. Alternatively, may be a list of additive covariate identifiers.

intcov Interactive covariates, entered in the same manner as addcov.

max.parents Maximum number of parents per node. This reduces the complexity of graphs and shortens run time. Probably best to consider values of 3-5.

parents List containing all possible parents up to max.parents in size. May be a subset

verbose Print iteration and number of models fit.

... Additional arguments passed to internal routines. In the case of bic.join, these are a list of objects produced by bic.qtlnet (see example below).

Details

The most expensive part of calculations is running scanone on each phenotype with parent phenotypes as covariates. One strategy is to pre-compute the BIC contributions using a cluster and save them for later use.

We divide the job into three steps: 1) determine parents and divide into reasonable sized groups; 2) compute BIC scores using scanone on a grid of computers; 3) compute multiple MCMC runs on a grid of computers. See the example for details.

```r
gr <- graph.qdg(acyclic.qdg, include.qtl = FALSE)
plot(gr)
```
bic.qtlnet

**Value**

Matrix with columns:

- **code**: Binary code as decimal for the parents of a phenotype node, excluding the phenotype. Value is between 0 (no parents) and $2^{\text{length(pheno.col)}-1}$.
- **pheno.col**: Phenotype column in reduced set, in range 1:length(pheno.col).
- **bic**: BIC score for phenotype conditional on parents (and covariates).

**Author(s)**

Brian S. Yandell and Elias Chaibub Neto

**References**


**See Also**

mcmc.qtlnet, parents.qtlnet

**Examples**

```r
pheno.col <- 1:13
max.parents <- 12
size.qtlnet(pheno.col, max.parents)

## Not run:
## Compute all phenotype/parent combinations.
## This shows how to break up into many smaller jobs.

##########################################################

pheno.col <- 1:13
max.parents <- 12
threshold <- 3.83

data(Pscdbp)
## or: load("Pscdbp.RData")
cross <- Pscdbp
cross <- read.cross("Pscdbp.csv", "csv")

## Break up into groups to run on several machines.
## ~53 groups of ~1000, for a total of 53248 scanone runs.
parents <- parents.qtlnet(pheno.col, max.parents)
groups <- group.qtlnet(parents = parents, group.size = 1000)

## Save all relevant objects for later steps.
```
save(cross, pheno.col, max.parents, threshold, parents, groups,
   file = "Step1.RData", compress = TRUE)

########################################################################
## STEP 2: Compute BIC scores. Parallelize.

## NB: Configuration of parallelization determined using Step 1 results.
## Load Step 1 computations.
load("Step1.RData")

## Parallelize this:
for(i in seq(nrow(groups)))
{
   ## Pre-compute BIC scores for selected parents.
   bic <- bic.qtlnet(cross, pheno.col, threshold,
      max.parents = max.parents,
      parents = parents[seq(groups[i,1], groups[i,2])])

   save(bic, file = paste("bic", i, ".RData", sep = ""), compress = TRUE)
}

########################################################################

## NB: n.runs sets the number of parallel runs.
n.runs <- 100

## Load Step 1 computations.
load("Step1.RData")

## Read in saved BIC scores and combine into one object.
bic.group <- list()
for(i in seq(nrow(groups)))
{
   load(paste("bic", i, ".RData", sep = ""))
   bic.group[[i]] <- bic
}
saved.scores <- bic.join(cross, pheno.col, bic.group)

## Parallelize this:
for(i in seq(n.runs))
{
   ## Run MCMC with randomized initial network.
   mcmc <- mcmc.qtlnet(cross, pheno.col, threshold = threshold,
      max.parents = max.parents, saved.scores = saved.scores, init.edges = NULL)

   save(mcmc, file = paste("mcmc", i, ".RData", sep = ""), compress = TRUE)
}

########################################################################
## STEP 4: Combine results for post-processing.
cyclica

## NB: n.runs needed from Step 3.
n.runs <- 100

## Combine outputs together.
outs.qtlnet <- list()
for(i in seq(n.runs))
{
  load(paste("mcmc", i, ".RData", sep = ""))
  outs.qtlnet[[i]] <- mcmc
}
out.qtlnet <- c.qtlnet(outs.qtlnet)
summary(out.qtlnet)
print(out.qtlnet)

## End of parallel example.
#####################################################

## End(Not run)
dim(Pscdbp.bic)

cyclica Cyclic graph (a) example

Description
We use a Gibbs sampling scheme to generate a data-set with 200 individuals (according with cyclic graph (a)). Each phenotype is affected by 3 QTLs. We fixed the regression coefficients at 0.5, error variances at 0.025 and the QTL effects at 0.2, 0.3 and 0.4 for the three F2 genotypes. We used a burn-in of 2000 for the Gibbs sampler.

Usage
data(cyclica)

Details
For cyclic graphs, the output of the qdg function computes the log-likelihood up to the normalization constant (un-normalized log-likelihood). We can use the un-normalized log-likelihood to compare cyclic graphs with reversed directions (they have the same normalization constant). However we cannot compare cyclic and acyclic graphs.

References

See Also
sim.cross, sim.geno, sim.map, skeleton, qdg, graph.qdg, generate.qtl.pheno
Examples

```r
## Not run:
bp <- matrix(0, 6, 6)
stdev <- rep(0.025, 6)

## Use R/qtl routines to simulate.
set.seed(3456789)
mymap <- sim.map(len = rep(100, 20), n.mar = 10, eq.spacing = FALSE,
    include.x = FALSE)
mycross <- sim.cross(map = mymap, n.ind = 200, type = "f2")
mycross <- sim.geno(mycross, n.draws = 1)
cyclicb.qtl <- generate.qtl.markers(cross = mycross, n.phe = 6)
mygeno <- pull.geno(mycross)[, unlist(cyclicb.qtl$markers)]
cyclicb.data <- generate.qtl.pheno("cyclicb", cross = mycross, burnin = 2000,
    bq = c(0.2, 0.3, 0.4), bp = bp, stdev = stdev, geno = mygeno)
save(cyclicb.qtl, cyclicb.data, file = "cyclicb.RData", compress = TRUE)

## End(Not run)
data(cyclicb)
out <- qdg(cross = cyclicb.data,
    phenotype.names = paste("y", 1:6, sep = ""),
    marker.names = cyclicb.qtl$markers,
    QTL = cyclicb.qtl$allqtl,
    alpha = 0.005,
    n.qdg.random.starts = 10,
    skel.method = "pcske1")

gr <- graph.qdg(out)
gr
plot(gr)
```

cyclicb  Cyclic graph (b) example

Description

We use a Gibbs sampling scheme to generate a data-set with 200 individuals (according with cyclic graph (b)). Each phenotype is affected by 3 QTLs. We fixed the regression coefficients at 0.5, error variances at 0.025 and the QTL effects at 0.2, 0.3 and 0.4 for the three F2 genotypes. We used a burn-in of 2000 for the Gibbs sampler.

Usage

data(cyclicb)
Details

For cyclic graphs, the output of the qdg function computes the log-likelihood up to the normalization constant (un-normalized log-likelihood). We can use the un-normalized log-likelihood to compare cyclic graphs with reversed directions (since they have the same normalization constant). However we cannot compare cyclic and acyclic graphs.

References


See Also

sim.cross, sim.geno, sim.map, skeleton, qdg, graph.qdg, generate.qtl.pheno

Examples

```r
## Not run:
bp <- matrix(0, 6, 6)
stdev <- rep(0.025, 6)

## Use R/qtl routines to simulate.
sset.seed(3456789)
mymap <- sim.map(len = rep(100,20), n.mar = 10, eq.spacing = FALSE,
                  include.x = FALSE)
mycross <- sim.cross(map = mymap, n.ind = 200, type = "f2")
mycross <- sim.geno(mycross, n.draws = 1)

cyclicb.qtl <- generate.qtl.markers(cross = mycross, n.phe = 6)
mygeno <- pull.geno(mycross)[, unlist(cyclicb.qtl$markers)]
cyclicb.data <- generate.qtl.pheno("cyclicb", cross = mycross, burnin = 2000,
                                    bq = c(0.2,0.3,0.4), bp = bp, stdev = stdev, geno = mygeno)
save(cyclicb.qtl, cyclicb.data, file = "cyclicb.RData", compress = TRUE)

## End(Not run)

data(cyclicb)
out <- qdg(cross=cyclicb.data,
          phenotype.names=paste("y",1:6,sep=""),
          marker.names=cyclicb.qtl$markers,
          QTL=cyclicb.qtl$allqtl,
          alpha=0.005,
          n.qdg.random.starts=10,
          skel.method="pcskel")

gr <- graph.qdg(out)
gr
plot(gr)
```
Description

We use a Gibbs sampling scheme to generate a data-set with 200 individuals (according with cyclic graph (c)). Each phenotype is affected by 3 QTLs. We fixed the regression coefficients at 0.5, (except for beta[5,2]=0.8) error variances at 0.025 and the QTL effects at 0.2, 0.3 and 0.4 for the three F2 genotypes. We used a burn-in of 2000 for the Gibbs sampler. This example illustrates that even though our method cannot detect reciprocal interactions (e.g. between phenotypes 2 and 5 in cyclic graph (c)), it can still infer the stronger direction, that is, the direction corresponding to the higher regression coefficient. Since beta[5,2] is greater than beta[2,5], the QDG method should infer the direction from 2 to 5.

Usage

data(cyclicc)

Details

For cyclic graphs, the output of the qdg function computes the log-likelihood up to the normalization constant (un-normalized log-likelihood). We can use the un-normalized log-likelihood to compare cyclic graphs with reversed directions (since they have the same normalization constant). However we cannot compare cyclic and acyclic graphs.

References


See Also

sim.cross, sim.geno, sim.map, skeleton, qdg, graph.qdg, generate.qtl.pheno

Examples

## Not run:
bp <- matrix(0, 6, 6)
bp[2,5] <- 0.5
bp[5,2] <- 0.8
stdev <- rep(0.025, 6)

## Use R/qtl routines to simulate map and genotypes.
set.seed(34567899)
mymap <- sim.map(len = rep(100,20), n.mar = 10, eq.spacing = FALSE, include.x = FALSE)
mycross <- sim.cross(map = mymap, n.ind = 200, type = "f2")
mycross <- sim.geno(mycross, n.draws = 1)
dist.qtlnet

## Use R/qdg routines to produce QTL sample and generate phenotypes.
cyclicc.qtlnet <- generate.qtl.markers(cross = mycross, n.phe = 6)
mygeno <- pull.geno(mycross)[, unlist(cyclicc.qtlnet$markers)]

cyclicc.data <- generate.qtl.pheno("cyclicc", cross = mycross, burnin = 2000,
  bq = c(0.2,0.3,0.4), bp = bp, stdev = stdev, geno = mygeno)
save(cyclicc.qtlnet, cyclicc.data, file = "cyclicc.RData", compress = TRUE)

## End(Not run)

data(cyclicc)
out <- qdg(cross = cyclicc.data,
phenotype.names = paste("y",1:6,sep=""),
marker.names = cyclicc.data$markers,
QTL = cyclicc.data$allqtl,
alpha = 0.005,
n.qdg.random.starts = 1,
skel.method = "pcskel")

gr <- graph.qdg(out)
plot(gr)

dist.qtlnet  QTL network diagnostic routines

Description

Various QTLnet diagnostic routines.

Usage

dist.qtlnet(qtlnet.object, min.prob = 0.9, method = "manhattan", cex = 5)
edgematch.qtlnet(qtlnet.object, min.prob = 0.9, method = "manhattan", cex = 5)
mds.qtlnet(qtlnet.object, min.prob = 0.9, method = "manhattan", cex = 5)
plotbic.qtlnet(x, ..., smooth = TRUE)

Arguments

qtlnet.object,x
  Object of class qtlnet.

min.prob
  Minimum probability to include edge in network.

method
  Distance method to be used between columns of connection matrix. Used by
dist. (Only used for mds.qtlnet.)

cex
  Character expansion. (Only used for mds.qtlnet, scaled by range of BIC val-
  ues.)

smooth
  Use lowess smoother if TRUE.

... Additional unused arguments.
generate.qtl

Value
List containing, for each phenotype in the network, a character vector of the QTL names as chr@pos, or pseudomarker name if chr.pos is FALSE.

Author(s)
Brian S. Yandell and Elias Chaibub Neto

References

See Also
mcmc.qtlnet

Examples
loci.qtlnet(Pscdbp.qtlnet)

---

generate.qtl Generate QTLs and phenotypes from cross object

Description
Generate QTLs and phenotype data for individual examples from cross. These are utility routines to illustrate the examples. They are not meant for users per se.

Usage
generate.qtl.markers(cross, n.phe, nqtl = 3)
generate.qtl.pheno(name, cross, bp, bq, stdev, allqtl, burnin = 2000, geno)

Arguments
cross object of class cross; see read.cross
name character string for example name
bp vector or matrix of coefficients for dependencies between phenotypes; see cyclic and acyclic examples
bq vector or matrix of coefficients for QTL effects on phenotypes; see cyclic and acyclic examples
stdev vector of standard deviations per phenotype
glxnet

allqtl  list of objects of class qtl produced by generate.qtl.sample
burnin  number of burnin cycles for MCMC; default is 2000
genopheno  genotypes at markers, typically extracted with pull.geno
n.phe  number of phenotypes
nqtl  number of QTL

See Also

acyclic, cyclica, cyclicb, cyclicc

Examples

## Not run:
example(acyclic)
example(cyclica)
example(cyclicb)
example(cyclicc)

## End(Not run)

---

glxnet  Generate and graph Glx network

Description

This is the Glx network reported in Chaibub Neto et al 2008 and in Ferrara et al 2008. Age was used as an additive covariate and we allowed for sex by genotype interaction. The network differs slightly from the published network due to improved code.

References


See Also

qdg
Examples

data(glxnet)
glxnet.cross <- calc.genoprob(glxnet.cross)
set.seed(1234)
glxnet.cross <- sim.geno(glxnet.cross)

n.node <- nphe(glxnet.cross) - 2  ## Last two are age and sex.
markers <- glxnet.qtl <- vector("list", n.node)
for(i in 1:n.node) {
  ac <- model.matrix(~ age + sex, glxnet.cross$pheno[, -1])
  ss <- summary(scanone(glxnet.cross, pheno.col = i,
                        addcovar = ac, intcovar = ac[,2]),
                threshold = 2.999)
  glxnet.qtl[[i]] <- makeqtl(glxnet.cross, chr = ss$chr, pos = ss$pos)
  markers[[i]] <- find.marker(glxnet.cross, chr = ss$chr, pos = ss$pos)
}
names(glxnet.qtl) <- names(markers) <- names(glxnet.cross$pheno)[seq(n.node)]

glxnet.qdg <- qdg(cross=glxnet.cross,
phenotype.names = names(glxnet.cross$pheno[,seq(n.node)]),
marker.names = markers,
QTL = glxnet.qtl,
alpha = 0.05,
n.qdg.random.starts=10,
addcov="age",
intcov="sex",
skel.method="udgskel",
udg.order=6)
glxnet.qdg
gr <- graph.qdg(glxnet.qdg)
plot(gr)
## Or use tkplot().
## Not run:
glxnet.cross <- clean(glxnet.cross)
save(glxnet.cross, glxnet.qdg, glxnet.qtl, file = "glxnet.RData", compress = TRUE)
## End(Not run)

igraph.qtlnet
qtlnet plot using igraph

Description

Plot inferred causal network using igraph package.

Usage

igraph.qtlnet(x, edges, loci.list,
   pheno.color = "green", qtl.color = "red", vertex.color,
include.qtl = TRUE, ...)
## S3 method for class 'qtlnet'
plot(x, ...)

**Arguments**

- **x** Object of class `qtlnet`.
- **edges** Data frame with first two columns being cause and effect directed phenotype pairs. Typically determined as averaged.net element from call to `summary.qtlnet`.
- **loci.list** List of character names of loci by phenotype. Typically determined by call to `loci.qtlnet`.
- **pheno.color, qtl.color** Name of color to use for phenotypes and QTLs, respectively.
- **vertex.color** Vertex colors in order of pheno-pheno edged augmented by loci.list. by default determined by pheno.color and qtl.color.
- **include.qtl** Include QTL in graph if TRUE and loci.list is not NULL.
- **...** Additional arguments passed to called routines.

**Details**

Uses the igraph package to create graph objects. These can be exported to a variety of other modern graphics packages. `graph.qtlnet` is synonymous with `igraph.qtlnet`.

**Value**

Object of class `graph` created using `graph.data.frame`.

**Author(s)**

Brian S. Yandell and Elias Chaibub Neto

**References**


**See Also**

`summary.qtlnet`, `loci.qtlnet`, `graph.data.frame`, `tkplot`

**Examples**

```r
pscdbp.graph <- igraph.qtlnet(Pscdbp.qtlnet)
pscdbp.graph
## Not run:
tkplot(Pscdbp.graph)

## End(Not run)
```
Description

Determines QTL that affect each phenotype conditional on the model-averaged network and on covariates.

Usage

loci.qtlnet(qtlnet.object, chr.pos = TRUE, merge.qtl = 10, ...)  
est.qtlnet(qtlnet.object, ..., verbose = TRUE)

Arguments

qtlnet.object Object of class qtlnet.
chr.pos Include chromosome and position if TRUE.
merge.qtl Merge QTL within merge.qtl cM of the mean QTL per chromosome across all nodes. No merge if 0 or less. This can reduce the number of QTL nodes to one per chr.
... Additional unused arguments.
verbose verbose output if TRUE.

Value

List containing, for each phenotype in the network, a character vector of the QTL names as chr@pos, or pseudomarker name if chr.pos is FALSE.

Author(s)

Brian S. Yandell and Elias Chaibub Neto

References


See Also

mcmc.qtlnet

Examples

loci.qtlnet(Pscdbp.qtlnet)
mcmc.qtlnet

Sample genetic architecture and QTL network

Description

Use MCMC to alternatively sample genetic architecture and QTL network as directed acyclic graphs (DAGs).

Usage

```r
mcmc.qtlnet(cross, pheno.col, threshold, addcov = NULL, intcov = NULL,
            nSamples = 1000, thinning = 1, max.parents = 3, M0 = NULL,
            burnin = 0.1, method = "hk", random.seed = NULL, init.edges = 0,
            saved.scores = NULL, rev.method = c("nbhd", "node.edge", "single"),
            verbose = FALSE, ...)
init.qtlnet(pheno.col, max.parents, init.edges)
```

Arguments

- **cross**: Object of class `cross`. See `read.cross`.
- **pheno.col**: Phenotype identifiers from `cross` object. May be numeric, logical or character.
- **threshold**: Scalar or list of thresholds, one per each node.
- **addcov**: Additive covariates for each phenotype (NULL if not used). If entered as scalar or vector (same format as `pheno.col`), then the same `addcov` is used for all phenotypes. Alternatively, may be a list of additive covariate identifiers.
- **intcov**: Interactive covariates, entered in the same manner as `addcov`.
- **nSamples**: Number of samples to record.
- **thinning**: Thinning rate. Number of MCMC samples is `nSamples*thinning`.
- **max.parents**: Maximum number of parents to a node. This reduces the complexity of graphs and shortens run time. Probably best to consider values of 3-5.
- **M0**: Matrix of 0s and 1s with initial directed graph of row->col if (row,col) entry is 1. Cycles are forbidden (e.g. 1s on diagonal or symmetric 1s across diagonal). Default (if NULL) is sampled by a call to `init.qtlnet`; all 0s if `init.edges` = 0 (default).
- **burnin**: Proportion of MCMC samples to use as burnin. Default is 0.1 if burnin is TRUE. Must be between 0 and 1.
- **method**: Model fitting method for `scanone`.
- **random.seed**: Initialization seed for random number generator. Must be NULL (no reset) or positive numeric. Used in `Random`.
- **init.edges**: Initial number of edges for `M0`, to be sampled using `{init.qtlnet}`. Chosen uniformly from 0 to the number of possible edges if set to NULL.
- **saved.scores**: Updated scores, typically pre-computed by `bic.qtlnet`.
- **rev.method**: Method to use for reversing edges. See details.
verbose Print iteration and number of models fit.
...
Additional arguments. Advanced users may want to supply pre-computed saved.scores to speed up calculations.

Details
Models are coded compactly as (1)(2|1)(3|1,2,4,5)(4|2)(5|2). Each parenthetical entry is a of form (node|parents); these each require a model fit, for now with scanone.

The scanone routine is run on multiple phenotypes in the network that could all have the same parents. For instance, for 5 phenotypes, if (1|2,4) is sampled, then do scanone of this model as well as (3|2,4) and (5|2,4). Setting the hidden parameter scan.parents to a value smaller than length(pheno.col) - 1 (default) disallows multiple trait scanning with more than that number of parents.

The saved.scores parameter can greatly reduce MCMC run time, by supplying pre-computed BIC scores. See bic.qtlnet. Another option is to capture saved.scores from a previous mcmc.qtlnet run with the same phenotypes (and covariates). Caution is advised as only a modest amount of checking can be done.

The init.qtlnet routine can be used to randomly find an initial causal network M0 with up to init.edges edges.

MCMC updates include delete, add or reverse edge direction. The early version of this method only considered the edge on its own (rev.method = "single"), while the neighborhood method (rev.method = "nbhd") uses the update

Value
List of class qtlnet

- post.model Model code (see details).
- post.bic Posterior BIC
- Mav Model average of M across MCMC samples.
- freq.accept Frequency of acceptance M-H proposals.
- saved.scores Saved LOD score for each phenotype and all possible sets of the other phenotypes as parent nodes.
- all.bic
- cross The cross object with calculated genotype probabilities.

In addition, a number of attributes are recorded:

- M0 Initial network matrix.
- threshold threshold list
- nSamples Number of samples saved
- thinning Thinning rate
- pheno.col Phenotype columns.
- pheno.names Phenotype names
- addcov Additive covariate columns.
intcov Interactive covariate columns.
burnin Burnin proportion
method Method used for scanone.
random.seed Initial random number generator seed.
random.kind Random number generator kind from Random.

Author(s)
Brian S. Yandell and Elias Chaibub Neto

References

See Also
read.cross, scanone, Random, bic.qtlnet.

Examples
data(Pscdbp)
## Not run:
## Run of subset of traits. Still takes some time.
Pscdbp.qtlnet <- mcmc.qtlnet(Pscdbp, pheno.col = c(1,2,4,5,6),
                           threshold = 3.83,
                           nSamples = 1000, thinning = 20,
                           random.seed = 92387475, verbose = TRUE)
save(Pscdbp.qtlnet, file = “Pscdbp.qtlnet.RData”, compress = TRUE)
## End(Not run)
data(Pscdbp.qtlnet)

## Not run:
out.qtlnet <- mcmc.qtlnet(Pscdbp, pheno.col = 1:13,
                          threshold = 3.83,
                          nSamples = 1000, thinning = 20,
                          random.seed = 92387475, verbose = TRUE,
                          saved.scores = Pscdbp.bic)
## End(Not run)
parallel qtlnet

Code to parallelize use of qtlnet

Description

This routine calls one of four phases in a parallelized version of qtlnet.

Usage

parallel.qtlnet(phase, index = 1, ..., dirpath = ")"

Arguments

phase  Phase of parallelization as number 1 through 4. See details.
index  Index for phase. Used in phases 2 and 4, and for error codes saved in RESULT.phase.index
       file.
...    Additional arguments for phases. See details.
dirpath Character string for directory where user can read and write files. When submitting to a cluster, this should remain the default.

Details

See http://www.stat.wisc.edu/~yandell/sysgen/qtlnet for details of implementation in progress. The plan is to run qtlnet via Condor (http://www.condor.org) to scale up to larger networks, say up to 100 nodes. Most important information is passed in files. Phase 1 imports arguments from the params.txt file, which must have parse-able assignments to the arguments of qtlnet::qtlnet.phase1. This first phase produces file Phase1.RData, which included objects used by all other phases.

Phase 1 also creates file groups.txt, which for each line has begin and end indices for the parents that would result from a call to parents.qtlnet. Phase 2 should be run the same number of times as the number of lines in file groups.txt. Each run produces a bicN.RData file containing BIC computations. These computations are aggregated in Phase 3 to create Phase3.RData, which contains the saved.scores used for mcmc.qtlnet runs in Phase 4, which each produce an mcmcn.RData file. The number of runs of Phase 4 is an argument nruns stored in the params.txt file processed in Phase 1. Finally, Phase 5 aggregates the MCMC results from multiple independent runs into one qtlnet object.

Author(s)

Brian S. Yandell and Elias Chaibub Neto

References


http://www.stat.wisc.edu/~yandell/sysgen/qtlnet
parents.qtlnet

See Also

mcmc.qtlnet, bic.qtlnet

Examples

## Not run:
parallel.qtlnet("/u/y/a/yandell/public/html/sysgen/qtlnet/condor", 1)

## Not run

---

**parents.qtlnet**  
*Determine and group node-parent combinations.*

**Description**

Routines useful for examining the size of node-parent combinations.

**Usage**

parents.qtlnet(pheno.col, max.parents = 3, codes.only = FALSE)

## S3 method for class 'parents.qtlnet'
summary(object, ...)

size.qtlnet(pheno.col, max.parents = 3)

group.qtlnet(pheno.col, max.parents = 3, n.groups = NULL,  
group.size = 50000, parents = parents.qtlnet(pheno.col, max.parents))

**Arguments**

- **pheno.col**: Phenotype identifiers from cross object. May be numeric, logical or character.
- **max.parents**: Maximum number of parents per node. This reduces the complexity of graphs and shortens run time. Probably best to consider values of 3-5.
- **parents**: List containing all possible parents up to max.parents in size. May be a subset.
- **codes.only**: Return only codes of parents if TRUE.
- **n.groups**: Number of groups for parallel computation. Determined from group.size if missing.
- **group.size**: Size of groups for parallel computation. See details.
- **object**: Object of class parent.qtlnet.
- **...**: Additional arguments ignored.

**Details**

The most expensive part of calculations is running scanone on each phenotype with parent phenotypes as covariates. One strategy is to pre-compute the BIC contributions using a cluster and save them for later use. The parents.qtlnet routine creates a list of all possible parent sets (up to max.parents in size). The size.qtlnet determines the number of scanone calculations possible for a network with nodes pheno.col and maximum parent size max.parents. The group.qtlnet groups the parent sets into roughly equal size groups for parallel computations. See **bic.qtlnet** for further details.
Value

The `size.qtlnet` returns the number of possible `scanone` computations needed for BIC scores.

The `group.qtlnet` produces and index into the parents list created by `parents.qtlnet`. See details.

The `parents.qtlnet` creates a list object with names being the code.

The summary method for such an object is a data frame with row names being the code, a binary code as decimal for the parents of a phenotype node, excluding the phenotype. Value is between 0 (no parents) and $2^{\text{length(pheno.col) - 1}}$. The columns are

- `parents`: Comma-separated string of parents to potential child node.
- `n.child`: Number of possible child nodes to this parent set.

Author(s)

Brian S. Yandell and Elias Chaibub Neto

References


See Also

`bic.qtlnet`

Examples

```r
## Restrict to at most 3 parents per node.
phenocol <- 1:6
max.parents <- 3
size.qtlnet(phenocol, max.parents)
pARENTS <- parents.qtlnet(phenocol, max.parents)
sUMMARY(pARENTS)

## Allow an arbitrary number (up to 12) of parents per node.
phenocol <- 1:13
max.parents <- 12
size.qtlnet(phenocol, max.parents)

## Make ~53 groups of ~1000, for a total of 53248 scanone runs.
pARENTS <- parents.qtlnet(phenocol, max.parents)
n.child <- sUMMARY(pARENTS)$n.child
tABLE(n.child)
gROUPS <- group.qtlnet(pARENTS = pARENTS, group.size = 1000)
apply(GROUPS, 1,
    function(group, pARENTS) sapply(pARENTS[seq(group[1], group[2])], length),
    pARENTS)
```
Cross and qtlnet objects with Ghazalpour et al. (2006) data. Only 13 phenotypes are included.

Description

The R/qtl cross object was created from data at source. The qtlnet object was created using mcmc.qtlnet.

Usage

data(Pscdbp)
data(Pscdbp.qtlnet)

Source

http://www.genetics.ucla.edu/labs/horvath/CoexpressionNetwork/MouseWeight

References


See Also

read.cross, mcmc.qtlnet

Examples

summary(Pscdbp)
## Not run:
summary(Pscdbp.qtlnet)

## End(Not run)
Description

This function implements the QDG algorithm described in Chaibub Neto et al 2008. It creates and scores QDGs. The computed scores (log-likelihood and BIC) are only valid for acyclic graphs. For cyclic networks qdgSEM should be used to compute the scores.

Usage

```r
qdg(cross, phenotype.names, marker.names, QTL, alpha,
   n.qdg.random.starts, addcov = NULL, intcov = NULL,
   skel.method = c("pcskel","udgskel"), udg.order = 2)
```

```
graph.qdg(x, ...)
## S3 method for class 'qdg'
print(x, ...)
## S3 method for class 'qdg'
summary(object, ...)
```

Arguments

- `cross`: object of class `cross` (see `read.cross`).
- `phenotype.names`: character string with names of phenotype nodes corresponding to phenotypes in `cross`.
- `marker.names`: list of character strings, one for each of `phenotype.names`. Each character string has the marker names for that phenotype.
- `QTL`: object of class `qtl` (see `makeqtl`).
- `alpha`: significance level threshold for PC or UDG algorithms (for the inference of the graph skeleton. See step 1 of the QDG algorithm). Must be between 0 and 1.
- `n.qdg.random.starts`: number of random starts for the QDG algorithm (see step 3 of the QDG algorithm).
- `addcov`: names of additive covariates. Must be valid phenotype names in `cross`. Expanded to include all `intcov` names.
- `intcov`: names of additive covariates. Must be valid phenotype names in `cross`.
- `skel.method`:Either "pcskel" for the PC skeleton algorithm (skeleton) or "udgskel" for the UDG algorithm (approximate.udg routine defined internal to qdg).
- `udg.order`: maximum allowed order of the UDG algorithm. Must be between zero and the number of variables minus 2.
- `x`: object of class `qdg`.
- `...`: additional arguments (ignored).
Details

The log-likelihood and BIC scores are computed based on the factorization of the joint distribution, and hence are only valid for acyclic networks. For cyclic networks these scores are relative to the unnormalized likelihoods. Models include phenotypes and QTLs. The 'udgskel' method for the computation of the skeleton of the causal model should be used for small networks only (the UDG algorithm quickly becomes computationally infeasible as the number of nodes increases).

Value

List object that inherits class "qdg" and "qdg" with components:

- **udg**: Undirected dependency graph from PC skeleton or UDG algorithms.
- **dg**: Directed dependency graph before recheck step (output of the step 2 of the QDG algorithm).
- **best.lm**: Solution with lowest BIC (best fit to the data).
- **solutions**: Solutions of dependency graph after recheck step (output of steps 3, 4 and 5 of the QDG algorithm.)
- **marker.names**: List of character strings, one for each of phenotype.names. Each character string has the marker names for that phenotype.
- **phenotype.names**: Character string with names of phenotype nodes corresponding to phenotypes in cross.

References


See Also

skeleton

Examples

```r
## simulate a genetic map (20 autosomes, 10 not equaly spaced markers per chromosome)
mymap <- sim.map(len=rep(100,20), n.mar=10, eq.spacing=FALSE, include.x=FALSE)

## simulate an F2 cross object with n.ind (number of individuals)
n.ind <- 200
mycross <- sim.cross(map=mymap, n.ind=n.ind, type="f2")

## produce multiple imputations of genotypes using the sim.geno function. The makeqtl function requires it,
## even though we are doing only one imputation (since we don't have missing data and we are using the
## genotypes in the markers, one imputation is enough)
ymcross <- sim.geno(mycross,n.draws=1)
```
## Sample markers (2 per phenotype)

```r
genotypes <- pull.geno(mycross)
geno.names <- dimnames(genotypes)[[2]]
m1 <- sample(geno.names, 2, replace=FALSE)
m2 <- sample(geno.names, 2, replace=FALSE)
m3 <- sample(geno.names, 2, replace=FALSE)
m4 <- sample(geno.names, 2, replace=FALSE)
```

## Get marker genotypes

```r
g11 <- genotypes[, m1[1]]; g12 <- genotypes[, m1[2]]
g12 <- genotypes[, m2[1]]; g22 <- genotypes[, m2[2]]
g31 <- genotypes[, m3[1]]; g32 <- genotypes[, m3[2]]
g41 <- genotypes[, m4[1]]; g42 <- genotypes[, m4[2]]
```

## Generate phenotypes

```r
y1 <- runif(3,0,5,1)[g11] + runif(3,0,5,1)[g12] + rnorm(n.ind)
y2 <- runif(3,0,5,1)[g21] + runif(3,0,5,1)[g22] + rnorm(n.ind)
y3 <- runif(1,0,5,1) * y1 + runif(1,0,5,1) * y2 + runif(3,0,5,1)[g31] + runif(3,0,5,1)[g32] + rnorm(n.ind)
y4 <- runif(1,0,5,1) * y3 + runif(3,0,5,1)[g41] + runif(3,0,5,1)[g42] + rnorm(n.ind)
```

## Incorporate phenotypes to cross object

```r
mycross$pheno <- data.frame(y1, y2, y3, y4)
```

## Create markers list

```r
markers <- list(m1, m2, m3, m4)
names(markers) <- c("y1", "y2", "y3", "y4")
```

## Create qtl object

```r
cross <- makeqtl(mycross, chr = m1.pos[, "chr"], pos = m1.pos[, "pos"])
cross <- makeqtl(mycross, chr = m2.pos[, "chr"], pos = m2.pos[, "pos"])
cross <- makeqtl(mycross, chr = m3.pos[, "chr"], pos = m3.pos[, "pos"])
cross <- makeqtl(mycross, chr = m4.pos[, "chr"], pos = m4.pos[, "pos"])
names(allqtl) <- c("y1", "y2", "y3", "y4")
```

## Infer QDG

```r
out <- qdg(cross=mycross, phenotype.names = c("y1", "y2", "y3", "y4"), marker.names = markers, QTL = allqtl, alpha = 0.005, n.qdg.random.starts=10, skel.method="pcskel")
```

```r
gr <- graph.qdg(out)
gr
```
Conduct permutation test for LOD score of edge direction on directed graph.

**Usage**

```r
qdg.perm.test(cross, nperm, node1, node2, common.cov = NULL,
DG, QTLs, addcov = NULL, intcov = NULL)
```

## S3 method for class 'qdg.perm.test'

- `summary(object, ...)`
- `print(x, ...)`

**Arguments**

- `cross`: Object of class `cross` (see `read.cross`).
- `nperm`: Number of permutations.
- `node1`: Character string with name of a phenotype node.
- `node2`: Character string with name of a phenotype node.
- `common.cov`: Character string with name of common phenotype covariates.
- `DG`: Directed graph of class `QDG`.
- `QTLs`: List of objects of class `qtl`.
- `addcov`: Names of additive covariates. Must be valid phenotype names in `cross`. Expanded to include all `intcov` names.
- `intcov`: Names of additive covariates. Must be valid phenotype names in `cross`.
- `x, object`: Object of class `qdg.perm.test`.
- `...`: Additional arguments ignored.

**Details**

`qdg.perm.test` performs `nperm` permutation-based test of LOD score for an edge of a directed graph.
Value

List composed by:

- `pvalue`  Permutation p-value.
- `obs.lod`  Observed LOD score.
- `PermSample`  Permutation LOD scores sample.
- `node1`  Character string with name of a phenotype nodes.
- `node2`  Character string with name of a phenotype nodes.

References


Examples

data(glxnet)
glxnet.cross <- calc.genoprob(glxnet.cross)
set.seed(1234)
glxnet.cross <- sim.geno(glxnet.cross)
## Should really use nperm = 1000 here.
qudg.perm.test(glxnet.cross, nperm = 10, "Glx", "Slc1a2",
               DG = glxnet.qdg$qdg, QTLs = glxnet.qtl)

qdg.sem  

Score directed graphs outputed by qdg using structural equation models (SEM)

Description

Score directed graphs (cyclic or acyclic) outputed by qdg function using the sem R package.

Usage

qdg.sem(qdgObject, cross)
## S3 method for class 'qdg.sem'
print(x, ...)
## S3 method for class 'qdg.sem'
summary(object, ...)

Arguments

- `qdgObject`  list containing the output of `qdg`.
- `cross`  object of class `cross` (see `read.cross`).
- `x,object`  object of class `qdg`.
- `...`  extra arguments to print or summary (ignored).
Details

Fits a SEM to the phenotypes network. QTLs are not included as variables in the model. When additive covariates are used in qdg, qdg.sem fits a SEM model to the residuals of the variables after adjustment of the additive covariates.

Value

List object that inherits class "qdg.sem" and "qdg" composed by:

- **best.SEM**: Solution with lowest SEM BIC (best fit to the data).
- **BIC.SEM**: Vector with the BIC values of all solutions from qdg.
- **path.coeffs**: Path coefficients associated with the best SEM solution.
- **Solutions**: Solutions of dependency graph after recheck step (output of steps 3, 4 and 5 of the QDG algorithm.)
- **marker.names**: List of character strings, one for each of phenotype.names. Each character string has the marker names for that phenotype.
- **phenotype.names**: Character string with names of phenotype nodes corresponding to phenotypes in cross.
- **dropped**: Indexes of solutions that were dropped (NULL if none dropped).

See Also

qdg sem

Examples

```r
# simulate a genetic map (20 autosomes, 10 not equally spaced markers per chromosome)
mymap <- sim.map(len=rep(100,20), n.mar=10, eq.spacing=FALSE, include.x=FALSE)

# simulate an F2 cross object with n.ind (number of individuals)
n.ind <- 200
mycross <- sim.cross(map=mymap, n.ind=n.ind, type="f2")

# produce multiple imputations of genotypes using the sim.geno function. The makeqtl function requires it, even though we are doing only one imputation (since we don't have missing data and we are using the genotypes in the markers, one imputation is enough)
mycross <- sim.geno(mycross, n.draws=1)

# sample markers (2 per phenotype)
genotypes <- pull.geno(mycross)
geno.names <- dimnames(genotypes)[[2]]
m1 <- sample(geno.names,2,replace=FALSE)
m2 <- sample(geno.names,2,replace=FALSE)
m3 <- sample(geno.names,2,replace=FALSE)
m4 <- sample(geno.names,2,replace=FALSE)
```
## get marker genotypes

g11 <- genotypes[,m1[1]]; g12 <- genotypes[,m1[2]]
g21 <- genotypes[,m2[1]]; g22 <- genotypes[,m2[2]]
g31 <- genotypes[,m3[1]]; g32 <- genotypes[,m3[2]]
g41 <- genotypes[,m4[1]]; g42 <- genotypes[,m4[2]]

## generate phenotypes

y1 <- runif(3,0.5,1)[g11] + runif(3,0.5,1)[g12] + rnorm(n.ind)
y2 <- runif(3,0.5,1)[g21] + runif(3,0.5,1)[g22] + rnorm(n.ind)
y3 <- runif(1,0.5,1) * y1 + runif(1,0.5,1) * y2 + runif(3,0.5,1)[g31] + runif(3,0.5,1)[g32] + rnorm(n.ind)
y4 <- runif(1,0.5,1) * y3 + runif(3,0.5,1)[g41] + runif(3,0.5,1)[g42] + rnorm(n.ind)

## incorporate phenotypes to cross object

mycross$pheno <- data.frame(y1,y2,y3,y4)

## create markers list

markers <- list(m1,m2,m3,m4)
names(markers) <- c("y1","y2","y3","y4")

## create qtl object

allqtls <- list()
m1.pos <- find.markerpos(mycross, m1)
allqtls[[1]] <- makeqtl(mycross, chr = m1.pos[,"chr"], pos = m1.pos[,"pos"])
m2.pos <- find.markerpos(mycross, m2)
allqtls[[2]] <- makeqtl(mycross, chr = m2.pos[,"chr"], pos = m2.pos[,"pos"])
m3.pos <- find.markerpos(mycross, m3)
allqtls[[3]] <- makeqtl(mycross, chr = m3.pos[,"chr"], pos = m3.pos[,"pos"])
m4.pos <- find.markerpos(mycross, m4)
allqtls[[4]] <- makeqtl(mycross, chr = m4.pos[,"chr"], pos = m4.pos[,"pos"])
names(allqtls) <- c("y1","y2","y3","y4")

## infer QDG

out <- qdg(cross=mycross, phenotype.names = c("y1","y2","y3","y4"), marker.names = markers, QTL = allqtls, alpha = 0.005, n.qdg.random.starts=10, skel.method="pcskel")
gr <- graph.qdg(out)
plot(gr)

## Following does not work. Not sure why.

out2 <- qdg.sem(out, cross=mycross)
out2
g2 <- graph.qdg(out2)
plot(g2)
subset.qtlnet

Catenate or subset qtlnet object(s).

Description

Multiple qtlnet objects can be catenated together or subsetted by run.

Usage

```r
## S3 method for class 'qtlnet'
subset(x, run, ...)
## S3 method for class 'qtlnet'
c(...)
best.qtlnet(x, burnin = attr(x, "burnin"), wh = which.min(meanbic(x, burnin)))
```

Arguments

- `x` Object of class qtlnet. See `mcmc.qtlnet`.
- `run` Numeric index to desired run. Must be between 0 and number of runs.
- `burnin` Proportion of MCMC samples to be considered as burnin. Taken from qtlnet object usually.
- `wh` Number identifying which model is best.
- `...` For `c.qtlnet`, objects of class qtlnet to be joined. Ignored for `subset.qtlnet`.

Details

The catenation is used by `parallel.qtlnet` in phase 5 to join together multiple independent MCMC runs. Note that the averaged network and the frequency of acceptance for a derived subset are only based on the saved samples, while the original qtlnet objects used all samples. Thus catenation and subset are not strictly reversible functions.

The `best.qtlnet` routine picks the run with the best (lowest) BIC score on average and returns that run as a qtlnet object. It also produces a trace plot of BIC for all the runs.

Value

Both return an object of class qtlnet.

Author(s)

Brian Yandell

See Also

`mcmc.qtlnet`
Examples

## Not run:
joined <- c(qtlnet1, qtlnet2)
sub1 <- subset(joined, 1)
best <- best.qtlnet(joined)
## qtlnet1 and sub1 should be nearly identical.
## End(Not run)

summary.qtlnet

summary of model average network and posterior table

Description

Print and summary methods for qtlnet objects.

Usage

## S3 method for class 'qtlnet'
print(x, cutoff = 0.01, digits = 3, ...)
## S3 method for class 'qtlnet'
summary(object, parent.patterns = FALSE, ...)
## S3 method for class 'summary.qtlnet'
print(x, ...)
check.qtlnet(object, min.prob = 0.9, correct = TRUE, verbose = FALSE, ...)

Arguments

x, object Object of class qtlnet.
cutoff Frequency cutoff for model patterns to be displayed. Always shows at least the most common pattern.
digits Number of digits to display for posterior probabilities on directed edges.
parent.patterns Include summary of parent patterns if TRUE.
min.prob Set the minimum posterior probability for inclusion of an edge.
correct Correct min.prob if TRUE.
verbose Print forbidden edges in model-averaged solution if TRUE.
... Other hidden arguments. These include min.prob, which can also be passed to other igraph.qtlnet and plot.qtlnet routines.

Author(s)

Brian S. Yandell and Elias Chaibub Neto
write.qtlnet

References

See Also
mcmc.qtlnet

Examples

data(Pscdbp.qtlnet)
print(Pscdbp.qtlnet)
summary(Pscdbp.qtlnet)

write.qtlnet(x, filename, edges, loci.list, include.qtl = TRUE, est.list, include.est = TRUE, digits = 3, col.names = TRUE, ...)

write.qtlnet as text file

Description
Write resulting graph as text file

Usage
write.qtlnet(x, filename, edges, loci.list, include.qtl = TRUE, est.list, include.est = TRUE, digits = 3, col.names = TRUE, ...)

Arguments
x
filename
edges
loci.list
include.qtl
est.list
include.est
digits
col.names
...
Details
Simple write of causal network, for instance to use with Cytoscape.

Value
Invisibly returns data frame that corresponds to saved file.

Author(s)
Brian S. Yandell and Elias Chaibub Neto

References

See Also
igraph.qtlnet

Examples
write.qtlnet(Pscdbp.qtlnet, "Pscdbp.txt")
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