Package ‘relSim’

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Author James M. Curran
Maintainer James M. Curran <j.curran@auckland.ac.nz>
Description A set of tools to explore the behaviour statistics used for forensic DNA interpretation when close relatives are involved. The package also offers some useful tools for exploring other forensic DNA situations.
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blockSim

Perform relatives simulations using large memory blocks in C

Description

Generate N pairs with a given relationship, calculate the LR for sibs, parent-child and the number of matching alleles and count the number of pairs that meet the threshold criteria.

Usage

```r
blockSim(N, Freqs, rel = "UN", ibsthresh = NULL, kithresh = NULL,
        code = 1, falseNeg = TRUE, BlockSize = N / 10)
```

Arguments

- **N**
  - The number of iterations to carry out

- **Freqs**
  - A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex.

- **rel**
  - generate unrelated (rel = 'UN'), full-sibs (rel = 'FS'), or parent child (rel = 'PC') pairs

- **ibsthresh**
  - A vector of one or more IBS thresholds

- **kithresh**
  - A vector of one or more KI/LR thresholds

- **code**
  - A code from 1 to 6 which dictates the events that will be counted.
    1. the LR for siblings will be compared to the values in kithresh and incremented if the LR is greater than the threshold
    2. the LR for parent/child will be compared to the values in kithresh and incremented if the LR is greater than the threshold
3. the number of matching alleles (IBS) will be compared to the values in ibsthresh and incremented if the IBS is greater than the threshold
4. the LR for siblings and the number of matching alleles will be compared to the values in kithresh and ibsthresh and incremented if both the LR and IBS is greater than the thresholds. ibsthresh and kithresh must be of equal length for this option to work
5. the LR for parent/child and the number of matching alleles will be compared to the values in kithresh and ibsthresh and incremented if both the LR and IBS is greater than the thresholds. ibsthresh and kithresh must be of equal length for this option to work
6. this option is equivalent to performing code 4 and 5 simultaneously. It is not currently implemented

falseNeg if TRUE then the number of results that DO NOT satisfy the conditions are counted, otherwise the number of results DO satisfy the conditions are counted

BlockSize Sets the number of random profiles to be generated in each iteration. By default the block size is set to 10 percent of the total sample size. It is unclear whether the procedure is more efficient if a bigger percentage of the total is used. Users must take care to make sure that the block size evenly divides N otherwise the procedure will exit. Users must also make sure that they have enough memory.

Details
This function is used for fast accurate estimation of false positive and false negative rates. It achieves part of its speed by block execution in C, and part by not saving the LR or IBS results. It can do 1 billion iterations in about an hour.

Value
A vector containing the number of profile pairs that satisfied the threshold conditions

Author(s)
James M. Curran

See Also
sim

Examples
## not run
## this counts the number of unrelated pairs that are falsely identified
## as siblings using the policy that there are 16 or more matching
## alleles, and the LR/KI is greater than 100,000
## this is a very rare event for the FBI Caucasians with a frequency of
## about 4-5 times in 10 million pairs
## Not run:
data(fbiCaucs)
N = 1e8
**Description**

This function simulates a population with an approximate level of population substructure. This is achieved by subdividing a population into equal sized subpopulations and allowing them to breed within themselves for

\[
t = \lceil \frac{\log_e (1 - \theta)}{\log \left(1 - \frac{1}{2N_s}\right)} \rceil
\]

generations, where \( N_s \) is the number of individuals in each subpopulation. This will produce a population with an estimated coancestry coefficient approximately equal to \( \theta \).

**Usage**

```r
breedFst(freqs, theta = 0.01, N = 10000, ns = 10, DNAtools = FALSE)
```

**Arguments**

- **Freqs** A list with an element, `freqs` which contains a list of vectors, where each vector is a set of allele frequencies for a locus
- **theta** A desired level of inbreeding, where \( 0 < \theta < 0.5 \)
- **N** Total population size
- **ns** The number of subpopulations. \( N/n_s \) needs to be greater than 100
- **DNAtools** If `TRUE` then the profiles in the return population will be formatted as a data frame with an id column and two columns per locus.

**Value**

An object of class 'population' which is a list with the following elements

- **profiles** a vector of profiles where the level of inbreeding is approximately equal to \( \theta \)
- **nProfiles** the total number of individuals in the population
- **nSubpops** the number of sub-populations in the population
- **nLoci** the number of loci each individual is typed at
- **theta** the desired level of substructure in the population. The actual value will be near to this.
- **Freqs** a Freq object representing the ancestral frequencies of the population
**calcFst**

**Author(s)**

James M. Curran

**Examples**

```r
data(uscaucs)
pop = breedFst(uscaucs)
```

**Description**

This procedure uses the method of Weir and Cockerham to estimate theta (Fst) for a population with substructure.

**Usage**

```r
calcFst(Pop, subPopIdx = NULL)
```

**Arguments**

- `Pop` An object type 'population'
- `subPopIdx` If this vector is not null, then it must consist of \( N \) elements with values from 1 to \( n_s \) representing which subpopulation each member of `Pop$profiles` belongs to. If it is null then it is assumed that the population consists of \( n_s \) subpopulations of equal size \( N_s \) so that \( n_s \times N_s = N \)

**Value**

A vector of length \( n_{loci} + 1 \) with locus-wise \( \theta \) values and an overall \( \theta \) value for the population.

**Author(s)**

James M. Curran

**References**


**See Also**

breedFst

**Examples**

```r
data(uscaucs)
p = breedFst(uscaucs)
fst = calcFst(p)
fst
```
**checkFreqs**  

*Make sure that the frequencies are such*

**Description**

Checks whether a list of frequencies at a series of genetic loci both sum to one and lie between 0 and 1.

**Usage**

`checkFreqs(Freqs)`

**Arguments**

- `Freqs`  
  A list containing elements `loci` and `freqs`. `freqs` is a list of vectors containing the frequencies at the given loci.

**Details**

If a locus fails to sum to one, or there are alleles which fall below zero or above one, then a warning message will be returned for each item in error.

**Author(s)**

James M. Curran

**See Also**

`normalizeFreqs`

**Examples**

```r
data(fbiCaucs)
checkFreqs(fbiCaucs)

## induce an error
fbCaucsfreqs[[1]] = runif(10)
checkFreqs(fbiCaucs)
```
errorRate

Returns the false positive or false negative rates for a set of IBS and/or KI thresholds

Description

This function is used to calculate the various tables in the work of Ge et al. and Balding et al. Specifically it can be used to calculate the false positive rate for unrelated pairs being identified as full-sibs or parent-child pairs under differing levels of IBS or KI (or both) thresholds. It can also be used to calculate the false negative rates for full-sib, or parent-child, pairs being identified as unrelated, again with differing levels of IBS, KI or both.

Usage

errorRate(simResults, bIBS = TRUE, bKI = FALSE,
          rel = "UN", IBStresh = 14:17,
          KItresh = c(1e3,1e4,1e5,1e6), nLoci = 13)

Arguments

simResults A data.frame with three columns labelled sib, pc and ibs. This will usually be obtained from a call to sim or readResults.

bIBS If TRUE then IBS thresholds are used to generate the error rates. If both bIBS and bKI are TRUE then both criteria are used.

bKI If TRUE then KI thresholds are used to generate the error rates. If both bIBS and bKI are TRUE then both criteria are used.

rel The relationship used in the simulation. Must be one of 'UN', 'FS' or 'PC'.

IBStresh A vector of IBS values that can be used to classify the results as being related (or not).

KItresh A vector of KI threshold values that can be used to classify the results as being related (or not).

nLoci The number of loci being used in the multiplex. This dictates the upper bound on the IBS values.

Value

A vector (or a two-column matrix) of false negative or false positive rates. If the relationship is 'UN' then false positive rates are returned for parent-child and full-sibs, with parent-child being in column 1 and full-sibs in column 2. If the relationship is 'PC' then the false negative rate is returned for parent-child pairs, and if it is 'FS' then the false negative rate for full-sibs.

Author(s)

James M. Curran
See Also

sim, readResults

Examples

```r
## not run
## Not run: data(fbiCaucs)
unrel = sim(10000)
errorRate(unrel)

## End(Not run)
```

---

**exclusionPower**

*Calculate the exclusion power of a multiplex by locus*

**Description**

Calculates the exclusion power

\[
1 - 2 \left( \sum_{i=1}^{n_1} p_i^2 \right)^2 - 4 \sum_{i=1}^{n_1} p_i^4
\]

at each locus for a set of allele frequencies.

**Usage**

`ep(Freqs)`

**Arguments**

- **Freqs**: A list containing two vectors and a list, called loci, counts, and freqs. The elements of loci are the loci present in the multiplex. The elements are freqs a vectors of allele frequencies for the locus. The elements of counts are irrelevant here.

**Value**

The exclusion power for each locus.

**Author(s)**

James M. Curran

**References**

Examples

data(fbiCaucs)
ep(fbiCaucs)

## get the multiplex wide exclusion power
1 - prod(1-ep(fbiCaucs))

---

**fbCaucs**

*CODIS STR Loci allele frequency data*

Description

This data structure

Usage

data(fbiCaucs)

Format

This data set is a list which has two sub-lists. The lists are named loci and freqs. loci is a vector of the 13 CODIS STR locus names. freqs is a list of 13 vectors, each vector contains the allele frequencies published for US Caucasians in Budowle et al. (2001).

Author(s)

James M. Curran

References


See Also

USCaucs

Examples

data(fbiCaucs)
names(fbiCaucs)
fbiCaucs$loci
names(fbiCaucs$freqs)
fbiCaucs$freqs[[1]]
names(fbiCaucs$freqs[[1]])
fbiCaucs$freqs[[1]][1]
fetchBMdata

Retrieves data from Budowle and Moretti (1999) from the web

Description
Retrieves the Budowle and Moretti (1999) and compiles the allele frequency tables needed for the other parts of this package such as sim.

Usage
fetchBMdata()

Details
The first three populations have data on 20 loci, the second three on 13 loci. The missing values (0’s in the raw data) have been dropped and are not used in calculating the frequencies. This function will not work if you are not connected to the internet, or access to the internet is blocked.

Value
A list consisting of six elements corresponding to the six populations detailed in the data set. Each of the list elements is a list in itself with two further elements named loci and freqs. loci is a vector of the 13-20 STR locus names. freqs is a list of 13-20 vectors, each vector contains the allele frequencies.

Author(s)
James M. Curran

References
Budowle, B. and Moretti, T.R. (1999), Genotype Profiles for Six Population Groups at the 13 CODIS Short Tandem Repeat Core Loci and Other PCR Based Loci, Forensic Science Communications 1(2).

See Also
fbiCaucs, USCaucs

Examples
## not run
## Not run:
db = fetchBMdata()
names(db)
f = db[["TRINIDADIAN"]]
freqs
dbExpect(f, k = "UN", collapse = TRUE)

## End(Not run)
**Identity by state**

**Description**

Calculates the total number of alleles that are shared by two profiles. If the two profiles in question are indeed relatives then the matching alleles may be identical by descent, or by random chance alone, hence identity by state.

**Usage**

\`IBS(prof1, prof2, nLoci = length(prof1)/2, bPrint = FALSE)\`

**Arguments**

- **prof1**: A matrix consisting of 2 columns and nLoci rows. Each entry in the matrix is the (coded) allele held by the individual.
- **prof2**: See prof1
- **nLoci**: The number of loci in the profiles. Specifying this value speeds up computation enormously.
- **bPrint**: If true then the result is printed locus by locus. This feature exists primarily for debugging purposes.

**Value**

An integer between 0 and 2*nLoci representing the total number of alleles that match in the two profiles.

**Author(s)**

James M. Curran

**Examples**

```r
data(fbiCaucs)
P1 = randomProfile(fbiCaucs)
C1 = randomChild(P1, fbiCaucs)
IBS(P1, C1)
IBS(P1, C1, bPrint = TRUE)
```
locusIBS  

Identity by state at a locus

Description

Calculates the number of alleles that are shared by two profiles at a single locus. If the two profiles in question are indeed relatives then the matching alleles may be identical by descent, or by random chance alone, hence identity by state.

Usage

locusIBS(profMat)

Arguments

profMat  
A matrix consisting of 4 columns and N rows. Each row in the matrix consists of the genotypes of two individuals.

Value

A vector of length N containing values 0, 1, or 2 depending on how many alleles each pair of profiles share at a locus.

Author(s)

James M. Curran

Examples

data(fbicaucs)
G = randomSample(1, fbicaucs, rel = 'FS', N = 1000)
ibs = locusIBS(G)
barplot(tabulate(ibs+1, nbins = 3))

lrMix  

Calculate locuswise likelihood ratios for two person victim/suspect mixtures

Description

Calculates the likelihood ratio for pairs of profiles under the propositions $H_p : \ V + S$ and $H_d : \ V + U$, where $V$, $S$ and $U$ are the victim, the suspect and someone unrelated to the suspect respectively. The calculation does not employ $\theta$ so there are no assumptions about the subpopulations of the contributors.
**Usage**

\[ \text{lrMix(profiles, Freqs)} \]

**Arguments**

- **profiles**: A vector of profile lists, from `randomProfilePairs`, `randomPCPairs` and `randomSibPairs` also work but should not really be used as the calculations do not take account of the relationship between the two individuals.

- **Freqs**: A list containing elements `freqs`, `loci` and `counts`. The element `freqs` is a list of vectors of allele frequencies at the loci listed in `loci`. These frequencies are used to evaluate the LR.

**Value**

A matrix of LRs calculated at each locus for every pair of profiles. Note this is the set of \( N \) profile pairs supplied in `profiles`, not a pairwise comparison.

**Author(s)**

James M. Curran

**Examples**

```r
data(USCaucs)
p = randomProfilePairs(USCaucs, 10000)
log.lrs = log10(lrmix(p, USCaucs))
boxplot(log.lrs, las = 2)
```

---

**lrPC**  
*Likelihood Ratio for Parent-Child / Paternity Index*

**Description**

Calculates Likelihood Ratio comparing the probability of two profiles if they are indeed parent-child compared to unrelated. This is the paternity index or PI.

**Usage**

```r
lrPC(parent, child, Freqs = NULL, 
nLocci = length(parent)/2, 
f = NULL, 
n = NULL)
```
Arguments

parent: A matrix consisting of 2 columns and nLoci rows. Each entry in the matrix is the (coded) allele held by the individual. This represents the alleged parent. The relationship is reflexive so it does not matter which profile is labelled parent and child.

child: See parent

Freqs: A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex. This argument or both f and n must be specified.

nLoci: The number of loci in the profiles

f: A concatenated vector of allele frequencies. Specifying this speeds up computation enormously.

n: A vector of length nLoci giving the number of alleles at each locus. Specifying this in advance enormously speeds up computation.

Value

A value between 0 and infinity representing support (or lack of support if the value is less than 1) for the hypothesis that the two profiles are parent and child. There is no mutation built into this calculation. This means that the LR will be zero if the profiles do not share at least one allele in common at each locus in the multiplex.

Author(s)

James M. Curran

References


See Also

lrSib, IBS

Examples

data(fbiCaucs)
P1 = randomProfile(fbiCaucs)
C1 = randomChild(P1, fbiCaucs)
lrPC(P1, C1, fbiCaucs)
lrSib

Likelihood Ratio / Kinship Index for full-siblings

Description

Calculates Likelihood Ratio comparing the probability of two profiles if they are indeed full-sibs compared to unrelated. This is sometimes called the kinship index (KI) for full-sibs.

Usage

lrSib(sib1, sib2, Freqs = NULL, nLoci = length(sib1)/2, f = NULL, n = NULL)

Arguments

sib1 A matrix consisting of 2 columns and nLoci rows. Each entry in the matrix is the (coded) allele held by the individual. This represents the alleged sibling. The relationship is reflexive so it does not matter which profile is labelled sib1 and sib2.

sib2 See sib1

Freqs A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex. This argument or both f and n must be specified

nLoci The number of loci in the profiles

f A concatenated vector of allele frequencies. Specifying this speeds up computation enormously

n A vector of length nLoci giving the number of alleles at each locus. Specifying this in advance enormously speeds up computation

Value

A value between 0 and infinity representing support (or lack of support if the value is less than 1) for the hypothesis that the two profiles are full-siblings. There is no mutation built into this calculation.

Author(s)

James M. Curran

References


See Also

lrSibDebug, lrPC, IBS
Examples

data(fbiCaucs)
P1 = randomProfile(fbiCaucs)
S1 = randomSib(P1, fbiCaucs)
P2 = randomProfile(fbiCaucs)
lrSib(P1, S1, fbiCaucs)
lrSib(P1, P2, fbiCaucs)

Description

Calculates Likelihood Ratio comparing the probability of two profiles if they are indeed full-sibs compared to unrelated. This is sometimes called the kinship index (KI) for full-sibs. This function is identical to lrSib except that the calculation is performed in R, and provides full calculation detail at each locus. It exists primarily to check that the correct formula and logic is being applied in the LR calculation so that the result can be manually verified.

Usage

lrSibDebug(sib1, sib2, Freqs)

Arguments

sib1 A matrix consisting of 2 columns and nLoci rows. Each entry in the matrix is the (coded) allele held by the individual. This represents the alleged sibling. The relationship is reflexive so it does not matter which profile is labelled sib1 and sib2.
sib2 See sib1
Freqs A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex.

Value

A list containing three elements Lines, lr, and Cases. Lines is a list of strings containing the calculation at each locus so that the result can be written to file for example. Cases is a numeric code listing which logical case (1-11) the locus falls into for the profiles in question. lr is the KI for full-sibs for the two profiles.

Author(s)

James M. Curran
normalizeFreqs

References

See Also
lrsib, lrPC, IBS

Examples
data(fbiCaucs)
P1 = randomProfile(fbiCaucs)
S1 = randomSib(P1, fbiCaucs)
P2 = randomProfile(fbiCaucs)
cat(paste(lrsibDebug(P1, S1, fbiCaucs)$Lines))
cat(paste(lrsibDebug(P1, P2, fbiCaucs)$Lines))

normalizeFreqs Normalize frequencies to 1

Description
Normalize a list of frequencies at a series of genetic loci both sum to one. Not that this does not deal with the problem of values larger than one or smaller than zero.

Usage
normalizeFreqs(Freqs)

Arguments
Freqs A list containing elements loci and freqs. freqs is a list of vectors containing the frequencies at the given loci.

Details
Divides vector in Freqs.freqs by the vector sum.

Value
A list containing elements loci and freqs. freqs is a list of vectors containing the frequencies at the given loci.

Author(s)
James M. Curran


See Also

checkFreqs

Examples

data(fbiCaucs)

## induce an error
fbCaucus$freqs[[1]] = rgamma(10,1,1)
checkFreqs(fbiCauc)

fbCaucus = normalizeFreqs(fbiCaucus)
checkFreqs(fbiCaucus)

print.population

Print summary details of a substructured population

Description

Nicely prints summary information about a substructured population created using breedFst

Usage

## S3 method for class 'population'
print(x, ...)

Arguments

x  The population object to be printed

...  Ignored - really should be passed to print, but given cat is actually called they are ignored

Author(s)

James M. Curran

See Also

breedFst

Examples

data(fbiCaucs)
p = breedFst(fbiCaucus)
print(p)
**print.profile**

**Print a DNA profile**

**Description**

Nicely prints a profile object out in genotype pairs

**Usage**

```r
## S3 method for class 'profile'
print(x, horizontal = FALSE, ...)
```

**Arguments**

- `x`: The profile object to be printed
- `horizontal`: if TRUE then the profile will print on a single line instead of multiple lines. Useful for comparing two profiles
- `...`: Ignored - really should be passed to print, but given cat is actually called they are ignored

**Author(s)**

James M. Curran

**Examples**

```r
data(fbiCaucs)
P1 = randomProfile(fbiCaucs)
P2 = randomProfile(fbiCaucs)
P1
print(P1, horizontal = TRUE)
print(P2, horizontal = TRUE)
```

---

**randomChild**

*Generate a random child from a given DNA profile and a given set of allele frequencies*

**Description**

Generates a random child (or parent) from a given DNA profile from a given set of allele frequencies. At each locus, the child inherits the first allele of the given profile with one half, or the second allele with probability one half. The second allele is chosen at random with probability proportional to the allele frequencies.
randomPCPairs

Usage

randomChild(profile, Freqs)

Arguments

profile  A vector of length 2*nLoci. Each entry in the vector is the (coded) allele held by the individual. This represents the parent. The relationship is reflexive so it does not matter if the profile is a parent or a child.

Freqs A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex.

Details

The alleles are simply integers rather than the STR repeat numbers. This speeds up computation immensely when calculating any of the LRs or IBS.

Value

A vector with 2*nLoci elements. Each pair of elements represents the genotype of the random individual at that locus. The genotype alleles are always ordered so that allele1 <= allele2.

Author(s)

James M. Curran

See Also

randomChild, randomSample, randomSib

Examples

data(fbiCaucs)
P1 = randomProfile(fbiCaucs)
C1 = randomChild(P1, fbiCaucs)
P1
C1

randomPCPairs  Generate one or more random parent/child pairs from a given set of allele frequencies

Description

Generates one or more pairs random parent/child pairs from a given set of allele frequencies.
randomProfile

Usage

randomPCPairs(Freqs, BlockSize = 1)

Arguments

Freqs A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex.

BlockSize The number of pairs of profiles to generate

Details

The alleles are simply integers rather than the STR repeat numbers. This speeds up computation immensely when calculating any of the LRs or IBS.

Value

A list of length BlockSize. Each element of the list has a sublist containing two profiles called parent and child

Author(s)

James M. Curran

See Also

randomSibPairs, randomProfilePairs

Examples

data(fbiCaucs)
P = randomPCPairs(fbiCaucs)
P$parent
nP$child

-----------

randomProfile Generate a random DNA profile from a given set of allele frequencies

Description

Generates a random DNA profile from a given set of allele frequencies.

Usage

randomProfile(Freqs)
Arguments

Freqs A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex.

Details

The alleles are simply integers rather than the STR repeat numbers. This speeds up computation immensely when calculating any of the LRs or IBS.

Value

A vector with 2*nLoci elements. Each pair of elements represents the genotype of the random individual at that locus. The genotype alleles are always ordered so that allele1 <= allele2.

Author(s)

James M. Curran

See Also

randomChild, randomSample, randomSib

Examples

data(fbiCaucs)
P1 = randomProfile(fbiCaucs)

randomProfilePairs Generate one or more random DNA profile pairs from a given set of allele frequencies

Description

Generates one or more random DNA profile pairs from a given set of allele frequencies.

Usage

randomProfilePairs(Freqs, BlockSize = 1)

Arguments

Freqs A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex.

BlockSize The number of pairs of profiles to generate
Details

The alleles are simply integers rather than the STR repeat numbers. This speeds up computation immensely when calculating any of the LRs or IBS.

Value

A list of length BlockSize. Each element of the list has a sublist containing two profiles called prof1 and prof2.

Author(s)

James M. Curran

See Also

randomPCPairs, randomSibPairs

Examples

data(fbicaucsI)
P = randomProfilePairs(fbicaucs)
P$prof1
P$prof2

randomSample Generate a random sample of related (or unrelated) pairs of people

Description

Generate a random sample of unrelated, full-sib, or parent/child pairs of profiles at a single locus.

Usage

randomSample(nLoc, Freqs, rel = "UN", N = 10000)

Arguments

nLoc The locus number to sample from
Freqs A list containing elements loci and freqs. freqs is a list of vectors containing the frequencies at the given loci.
rel One of 'UN', 'FS', or 'PC' for unrelated, full-sib, or parent/child pairs respectively.
N The sample size
randomSib

Value
An N by 4 matrix of random profiles. The first two columns represent the genotype of person one
and the second two columns represent the genotype of column two. Note that the random profiles
do not use the orginal allele designations.

Author(s)
James M. Curran

See Also
randomProfile, randomSib, randomChild

Examples
```
data(fbicaucs)
G = randomSample(1, fbicaucs, "FS", 100)
```

randomSib

Generate a random sibling from a given DNA profile and a given set
of allele frequencies

Description
Generates a random sibling from a given DNA profile from a given set of allele frequencies. At each
locus, the sibling inherits the first allele of the given profile with one quarter, or the second allele
with probability one quarter, both alleles with probability one quarter, or neither with probability
one quarter. If the sibling inherits zero or one identical alleles, the missing alleles are chosen at
random with probability proportional to the allele frequencies.

Usage
```
randomSib(profile, Freqs)
```

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>profile</td>
<td>A vector consisting of 2*nLoci elements. Each element in the vector is the (coded) allele held by the individual. This represents the sibling.</td>
</tr>
<tr>
<td>Freqs</td>
<td>A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex.</td>
</tr>
</tbody>
</table>

Details
The alleles are simply integers rather than the STR repeat numbers. This speeds up computation
immensely when calculating any of the LRs or IBS.
randomSibPairs

Value
A vector with 2*nLoci elements. Each pair of elements represents the genotype of the random individual at that locus. The genotype alleles are always ordered so that allele1 <= allele2.

Author(s)
James M. Curran

See Also
randomChild, randomSample

Examples
data(fbiCaucs)
P1 = randomProfile(fbiCaucs)
S1 = randomSib(P1, fbiCaucs)
P1
S1

Description
Generates one or more pairs of random siblings from a given set of allele frequencies.

Usage
randomSibPairs(Freqs, BlockSize = 1)

Arguments
Freqs A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex.

BlockSize The number of pairs of profiles to generate

Details
The alleles are simply integers rather than the STR repeat numbers. This speeds up computation immensely when calculating any of the LRs or IBS.

Value
A list of length BlockSize. Each element of the list has a sublist containing two profiles called sib1 and sib2
Author(s)

James M. Curran

See Also

randomPCPairs, randomProfilePairs

Examples

data(fbicaucs)
P = randomSibPairs(fbicaucs)
P$sib1
P$sib2

---

readFreqs  Read in a file of allele frequencies

Description

Reads in a file of alleles in a particular format.

Usage

readFreqs(strPath, FSIGenFormat = TRUE, delim = ',')

Arguments

strPath  The file from which to read the frequencies
FSIGenFormat  Tells the function whether the file is either in FSI Genetics format (see below) or 'Curran' format
delim  This argument is used when FSIGenFormat is TRUE, and is the regular expression used to delimit columns of the table. It is set to a single comma by default, and multiple delimiters are considered empty separate fields. There probably should be an additional argument which specifies the missing or empty cell symbol, but I won't programme this unless somebody asks for it

Details

This function reads frequencies in the rectangular allele frequency table format used by FSI Genetics and other journals. This file format assumes a comma separated value file (CSV) (although the column delimiter can be specified). The first column should be labelled 'Allele' and contain the STR allele designations that are used in the data set. The remaining columns will have the locus name as a header, and frequencies that are either blank, zero, or non-zero. Blanks or zeros are used to specify that the allele is not observed (and not used) at the locus. The final row of the file should start with 'N' or 'n' in the first column and give the number of individuals typed (or the number of alleles recorded) in assessing the frequency of the alleles.
The second format is a very particular 'Curran' text format. The first line contains the number of loci in the multiplex. The next line will contain the name of the first locus and the number of alleles, \( nA \), the locus separated by a comma. The next \( nA \) lines contain the allele number (from 1 to \( nA \)), the STR designation of the allele, and the frequency separated by commas. This pattern is repeated for each locus. In the future this function will read the rectangular allele frequency table used by FSI Genetics and other journals.

Value

a list containing two vectors and a list, loci, counts, and freqs. The vector loci is a vector of the locus names in the frequency file. The vector counts is a vector of the number of individuals (or sometimes alleles) typed at each locus. This will null if the 'Curran' format is used. The list freqs, is a list of vectors with each vector containing the frequencies of the alleles at the locus. The names of the elements of the vectors are the STR allele designations.

Author(s)

James M. Curran

readResults

Read a simulation result set from file

Description

This function will read the output from sim that has been saved to disk

Usage

readResults(N = 0, rel = "UN", gzip = TRUE, strPath = "", strVer = "",
fileName = NULL)

Arguments

- **N**: The number of iterations in the simulation
- **rel**: 'UN' = unrelated, 'FS' = full-sib, 'PC' = parent-child
- **gzip**: If TRUE then it is assumed that the file is compressed
- **strPath**: Optional location of files. Must terminate with / otherwise it will not work
- **strVer**: A version string, useful if more than simulation has been run
- **fileName**: This argument allows the user to override the default file naming conventions of the result file

Details

The arguments to this file are used to generate the input file name. The format is very rigid, being 'results-sim-rel-N(-strVer).csv(.gz)' That is, if strVer is something than an empty string then it is included after the number of iterations. Similarly if gzip == TRUE then the filename is assumed to end with '.gz'
Value

A data frame with three columns labelled sib, pc, and ibs. These represent the LRs for sibs and parent-child calculated on each simulated profile pair, and the number of matching alleles (IBS).

Author(s)

James M. Curran

See Also

sim

Examples

data(fbiCaucs)
## not run
## write the results of 100 unrelated profile pairs to
## results-sim-UN-100.csv.gz
## and read it back in
## Not run:
sim(100, save = T)
unrel = readResults(100)
sim(100, rel = "FS", strVer = "01", save = T)
sibs = readResults(100, rel = "FS", strVer = "01")

## End(Not run)

---

**sim**

*Perform the relatives simulation*

Description

Generate N pairs with a given relationship and calculate the LR for sibs, parent-child and the number of matching alleles

Usage

```r
sim(N, Freqs, rel = "UN", save = FALSE, strPath = "", strVer = ",
BlockSize = N/100, fileName = NULL)
```

Arguments

- **N**
  - The number of iterations to carry out
- **Freqs**
  - A list containing two lists labelled loci and freqs. The second list is a list of vectors containing the allele frequencies of each allele at each locus in the multiplex.
- **rel**
  - generate unrelated (rel = 'UN'), full-sibs (rel = 'FS'), or parent child (rel = 'PC') pairs
save Write the results to disk if TRUE
strPath Optional prefix to add to the results file path so that the output location can be specified
strVer Optional suffix for the results file. This is useful when running multiple instances of R
BlockSize Sets the number of random profiles to be generated in each iteration. By default the block size is set to 1 percent of the total sample size. It is unclear whether the procedure is more efficient if a bigger percentage of the total is used. Users must take care to make sure that the block size evenly divides N otherwise the procedure will exit
fileName This argument lets the user override the default result file naming scheme

Details

This is the function that generates all the data for the results in the paper. WARNING: this function is not especially fast. To achieve the 100 million iterations used in the paper, 30 instances of R were launched on a multicore server. Each instance represented one relationship with 10 million iterations. The compute time for this arrangement was approximately 1 hours, meaning a full serial run would have taken over 30 hours to achieve the same result.

Value

a data frame with three columns: sib, pc, ibs containing the LRs for full-siblings, parent-child, and the number of matching alleles for each generated pair of profiles.

Author(s)

James M. Curran

See Also

readResults, errorRate

Examples

```r
## not run
## this replicates Ge et al.'s experiment and takes about 45 minutes
## to run (I think)
## Not run:
data(fbiCaucs)
N = 1000000
sim(N, fbiCaucs, save = T)
sim(N, fbiCaucs, 'FS', save = T)
sim(N, fbiCaucs, 'PC', save = T)
## End(Not run)
```
Exports a population with population substructure to a Nexus formatted file so that GDA can be used to check the Fst calculations.

Usage:
```
> tonexus(Pop, fileName = 'output.nex')
```

Arguments:
- `Pop`: An object of type 'population' - see `breedFst` for a description of the object.
- `fileName`: The name of the file output file.

Author(s):
James M. Curran

References:


See Also:
- `breedFst`

Examples:
```
data(USCaucs)
> p = breedFst(USCaucs)
> tonexus(p)
```
Description
This data structure

Usage
data(USCaucs)

Format
This data set is a list which has two sub-lists. The lists are named loci and freqs. loci is a vector of the 13 CODIS STR locus names. freqs is a list of 13 vectors, each vector contains the allele frequencies published for US Caucasians in Budowle and Moretti (1999). The raw data is available from http://www.fbi.gov/about-us/lab/forensic-science-communications/fsc/july1999/dnaloci.txt

Author(s)
James M. Curran

References
Budowle, B. and Moretti, T.R. (1999), Genotype Profiles for Six Population Groups at the 13 CODIS Short Tandem Repeat Core Loci and Other PCR Based Loci, Forensic Science Communications 1(2).

See Also
fbiCaucs

Examples
data(USCaucs)
names(USCaucs)
USCaucs$loci
names(USCaucs$freqs)
USCaucs$freqs[[1]]
names(USCaucs$freqs[[1]])
USCaucs$freqs[[1]][1]
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